








Probing cardiac metabolism by hyperpolarized ^{13}C MR using an exclusively endogenous substrate mixture and photo-induced nonpersistent radicals

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Purpose: To probe the cardiac metabolism of carbohydrates and short chain fatty acids simultaneously *in vivo* following the injection of a hyperpolarized ^{13}C -labeled substrate mixture prepared using photo-induced nonpersistent radicals.

Methods: Droplets of mixed $[1-^{13}\text{C}]$ pyruvic and $[1-^{13}\text{C}]$ butyric acids were frozen into glassy beads in liquid nitrogen. Ethanol addition was investigated as a means to increase the polarization level. The beads were irradiated with ultraviolet light and the radical concentration was measured by ESR spectroscopy. Following dynamic nuclear polarization in a 7T polarizer, the beads were dissolved, and the radical-free hyperpolarized solution was rapidly transferred into an injection pump located inside a 9.4T scanner. The hyperpolarized solution was injected in healthy rats to measure cardiac metabolism *in vivo*.

Results: Ultraviolet irradiation created nonpersistent radicals in a mixture containing ^{13}C -labeled pyruvic and butyric acids, and enabled the hyperpolarization of both substrates by dynamic nuclear polarization. Ethanol addition increased the radical concentration from 16 to 26 mM. Liquid-state ^{13}C polarization was 3% inside the pump at the time of injection, and increased to 5% by addition of ethanol to the substrate mixture prior to ultraviolet irradiation. In the rat heart, the *in vivo* ^{13}C signals from lactate, alanine, bicarbonate, and acetylcarnitine were detected following the metabolism of the injected substrate mixture.

Conclusion: Copolarization of two different ^{13}C -labeled substrates and the detection of their myocardial metabolism *in vivo* was achieved without using persistent radicals. The absence of radicals in the solution containing the hyperpolarized ^{13}C -substrates may simplify the translation to clinical use, as no radical filtration is required prior to injection.

KEYWORDS

carbon-13, energy metabolism, hyperpolarization, metabolic imaging, oxidative metabolism

1 | INTRODUCTION

Hyperpolarization methods overcome the relatively low sensitivity of MRS, and by combining dissolution dynamic nuclear polarization (DNP)¹ and ¹³C MRS imaging, *in vivo* metabolism can be probed in real time.² Hyperpolarized ¹³C MR has already been used extensively in a wide range of preclinical models, in which it was shown to be sensitive to variations in key metabolic pathways.^{3–8} The technique has recently been translated to studies investigating prostate cancer and heart disease in human subjects.^{9,10} Further clinical applications in cardiology and oncology have been discussed,^{11,12} and several more sites around the world are now poised to initiate clinical trials.

Because of the heart's high substrate uptake rate, cardiac applications are well adapted to the relatively short life time of the hyperpolarized ¹³C MR signals. Heart metabolism is flexible, exhibiting the ability to switch between the oxidation of different substrates, depending on their availability and metabolic state, but substrate preference is often disturbed and shifted in disease.¹³ Using hyperpolarized ¹³C MR, it was recently shown that both short chain fatty acid and carbohydrate oxidation can be simultaneously monitored *in vivo*, in real time, and in a single experiment, following the co-administration of 2 different substrates.¹⁴ However, this promising substrate mixture has not yet been validated for clinical applications, and the initial preclinical experiments were done using nitroxyl radicals.

To hyperpolarize the ¹³C spins by means of DNP, it is necessary to admix persistent radicals with the ¹³C-labeled biomolecules of interest.¹ These radicals must be removed prior to injection into humans, and the polarization of pyruvic acid for clinical applications is currently performed with a specific form of costly trityl radical that can be extracted from an acidic solution.¹⁵ The inline filtration process might be more challenging, however, for complex mixtures of biomolecules. For regulatory reasons, it is also necessary to add a quality control test to ensure that the filtration process was efficient and that the residual concentration of radical is below an acceptable value, a step that adds complexity to the process, possibly delaying the release of the hyperpolarized ¹³C substrates for injection, and that can potentially fail.

It was recently demonstrated that it is possible to perform DNP using nonpersistent radicals induced by ultraviolet (UV) irradiation of pyruvic acid.¹⁶ These radicals can be used to efficiently hyperpolarize [1-¹³C]pyruvic acid as well as other ¹³C-biomolecules and isotopes with nuclear spin.^{16,17} The radicals disappear during dissolution, recombining to form ¹³CO₂ and unlabeled acetate.¹⁶ As a result, the hyperpolarized solution can be formulated to contain only endogenous substrates. The use of photo-induced

nonpersistent radicals may also reduce experimental costs, as they alleviate the need for synthetic persistent radicals. Another promising development recently demonstrated that photo-induced radicals can be annihilated without dissolving the frozen substrates, allowing storage and transport of hyperpolarized substances.¹⁸ This may lead to a paradigm shift in DNP, providing applications for sites that are not equipped with their own DNP equipment to perform clinical studies.

The purpose of the present work was to investigate whether photo-induced nonpersistent radicals could be used to copolarize a mixture of ¹³C-substrates for the simultaneous measurement of separate biochemical pathways *in vivo*.

2 | METHODS

2.1 | Sample preparation

All chemicals were purchased from Sigma-Aldrich (Buchs, Switzerland). A solution consisting of 50 μ L [1-¹³C]pyruvic acid and 50 μ L [1-¹³C]butyric acid was pipetted dropwise in liquid nitrogen to form frozen glassy beads of approximately 10 μ L (diameter of approximately 2.5 mm), which were collected inside the 6-mm inner-diameter tail of a quartz dewar designed for ESR measurements (Wilmad LabGlass 150-mL Suprasil Dewar Flask type WG-850-B-Q, Vineland, NJ). The frozen beads were then irradiated with UV light for a total of 1 hour using a 365-nm LED array (Hamamatsu Photonics LC-L5, Hamamatsu City, Japan) following a previously described procedure.¹⁷ This process creates the radicals necessary for DNP. After completion of the irradiation, the beads were transferred to a glass vial for storage in liquid nitrogen. It was also investigated whether radical concentration and polarization level can be increased by modifying the substrate mixture. Therefore, a second solution consisting of [1-¹³C]pyruvic acid, [1-¹³C]butyric acid, and ethanol in a 2:2:1 ratio (v/v) was prepared separately. The mixture containing ethanol, however, was not used for *in vivo* experiments.

2.2 | Radical concentration measurements

In a separate set of experiments, the radical concentration was measured as a function of the UV irradiation time using an X-band ESR spectrometer (Bruker EMX, Billerica, MA). Measurements were performed at 77 K on frozen UV-irradiated beads containing only [1-¹³C]pyruvic and [1-¹³C]butyric acid, as well as in UV-irradiated beads containing an added 20% volume of ethanol. The ESR measurements of the UV-irradiated mixtures were performed at 10-minute intervals during the 1-hour irradiation process, and the radical concentration was plotted as a function of the total irradiation time.

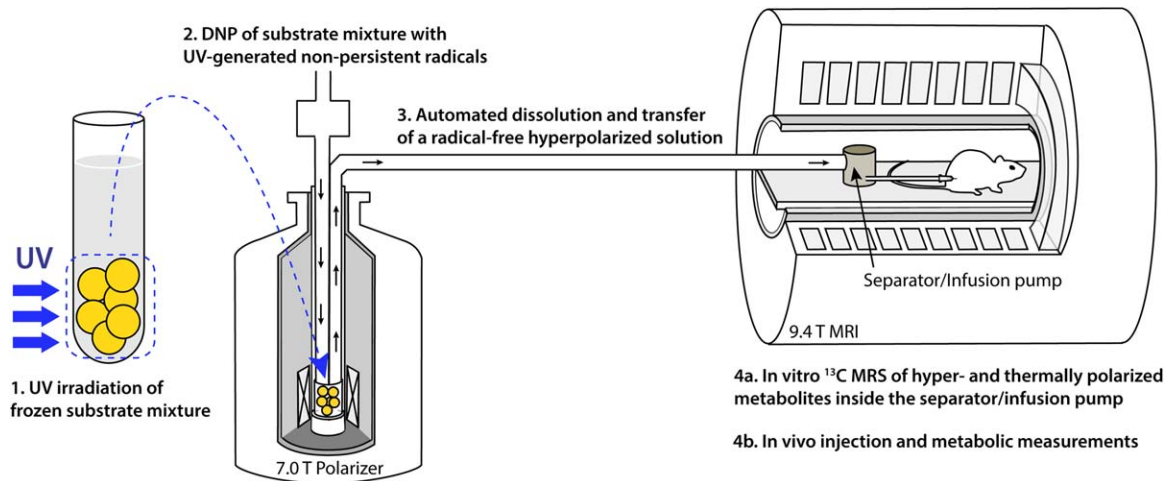


FIGURE 1 Illustration of the in vivo and in vitro experimental method based on ultraviolet (UV) irradiated ^{13}C -labeled metabolic substrate mixtures: (1) a frozen mixture of ^{13}C -labeled substrates, in this case [^{13}C]butyric and [^{13}C]pyruvic acids, is irradiated with UV light at 77 K; (2) the mixture is loaded into a DNP polarizer and ^{13}C nuclei are dynamically polarized for 1 to 2 hours; (3) using an automated process, the sample is quickly dissolved in superheated buffer solution and automatically transferred from the polarizer into a separator/injection pump located inside the bore of an MRI magnet; and (4) in vitro ^{13}C MRS measurements are performed while the hyperpolarized ^{13}C -substrate mixture is inside the separator/injection pump and/or the mixture is injected into the rat via a femoral vein catheter, and in vivo hyperpolarized ^{13}C MRS measurements are launched

2.3 | Polarization and dissolution

The frozen glassy UV-irradiated beads were loaded into a polytetrafluoroethylene sample cup together with frozen droplets of NaOH solution, the volume and concentration of which was calculated to balance the pH to 7 during dissolution. The polytetrafluoroethylene cup was then placed inside a 7T homebuilt polarizer, and the ^{13}C spins were polarized for 2 hours at 1.0 ± 0.1 K with the microwave frequency set to 196.75 GHz.^{19–21} Using an automated process,²² the sample was rapidly dissolved in 6 mL D_2O and transferred within 2 seconds into a remotely controlled phase separator/injection pump,²³ located inside the bore of the MR scanner (Figure 1). For the in vivo experiments, the separator/injection pump was prefilled with 0.6 mL of phosphate-buffered saline and heparin, and 800 μL of the hyperpolarized ^{13}C -labeled substrate mixture was automatically injected into the femoral vein of the animal.

2.4 | In vitro hyperpolarized and thermal ^{13}C MRS

In vitro experiments were performed to determine the ^{13}C signal enhancement and polarization levels for both types of UV-irradiated mixtures after hyperpolarization and transfer into the separator/injection pump. All in vitro ^{13}C MRS measurements were performed within the pump as previously described²² using a custom-made dual $^1\text{H}/^{13}\text{C}$ probe wrapped around the body of the pump. The hyperpolarized ^{13}C MRS acquisition was triggered at the start of the infusion process, collecting transient spectra with a 3-second

repetition time and RF excitation angle of 5° . Two different quantitative methods were used to measure the thermal ^{13}C polarization to doubly verify the polarization levels obtained after dissolution, as each method may be subject to different potential sources of error (see Supporting Information). In the first method, the ^{13}C signal was measured using 90° RF excitation pulses with a repetition time of 240 seconds and 8 averages, as described previously.²⁴ The second method consisted of measuring the ^{13}C signal with a 5° RF excitation angle, repetition time of 1.1 seconds, and 1024 averages following the addition of 5 μL of gadolinium complex (1 mmol/mL, Dotarem, gadoteric acid, 0.5M, Guerbet, Villepinte, France) to a concentration of approximately 1 mM, as described previously.²¹

2.5 | Animals

All animal experiments were conducted according to federal ethical guidelines and were approved by the local regulatory body. Male Sprague Dawley rats were initially anesthetized with 5% isoflurane in oxygen. Catheters were placed into the femoral vein for substrate delivery and in the femoral artery to monitor the blood pressure and heart rate, as previously described.^{25,26} The respiration rate, cardiac rhythm, and temperature were monitored and maintained using a pneumatic respiration sensor, a blood pressure sensor, and a rectal temperature probe (Small Animal Instruments Inc, Stony Brook, NY), respectively. After surgery, the isoflurane concentration was decreased to 1.5% in oxygen. Animals were placed supine in a custom-designed animal holder, and heating was provided by warm water circulating through tubing placed next to the rat.

2.6 | In vivo MRI and hyperpolarized ^{13}C MRS

All MRI and MRS measurements were carried out in a horizontal bore 9.4T magnet (Magnex Scientific, Oxford, United Kingdom) with a Direct Drive spectrometer (Varian, Palo Alto, CA). A custom-made RF hybrid probe, consisting of a 10-mm-diameter proton surface coil and a pair of 10-mm-diameter ^{13}C surface coils in quadrature mode, was positioned over the chest of the rat for transmission and reception. A hollow glass sphere with a 3-mm inner diameter (Wilmad-LabGlass) was filled with an aqueous 1 M [$1\text{-}^{13}\text{C}$] glucose solution and used to adjust the RF excitation pulse power and set the reference frequency. Acquisition of gradient echo proton images confirmed the correct placement of the coil and was used to determine the voxel used for shimming.

Cinematographic images ($\text{FOV} = 40 \times 40 \text{ mm}^2$; matrix size: 256×256 ; $\text{TR} = 140 \text{ ms}$; $\text{TE} = 4.5 \text{ ms}$; number of acquisitions = 8; number of frames = 14; slice thickness = 1 mm) were acquired to confirm and set the timing of the cardiac trigger in the end-diastolic phase. The cardiac trigger was typically sent 50 or 60 ms after the observed maximum blood pressure. Cardiac-triggered and respiratory-gated shimming was performed using the FAST(EST)MAP gradient shimming routine²⁷ to reduce the localized proton line width in a myocardial voxel of $4 \times 5 \times 5 \text{ mm}^3$ (acquired by stimulated echo localized spectroscopy) to 20 to 30 Hz, resulting in a nonlocalized proton line width of 80 to 120 Hz. The MR console was triggered to start acquisition at the beginning of the automated injection process of the hyperpolarized ^{13}C substrate mixture. A series of single-pulse acquisitions were sequentially recorded using 30° adiabatic RF excitation pulses (BIR-4),²⁸ with ^1H decoupling using WALTZ.²⁹ Free induction decays were acquired with 4129 complex data points over a 20-kHz bandwidth. All acquisitions were cardiac-triggered and respiratory-gated, resulting in a TR between 3 and 3.5 seconds. The adiabatic pulse offset and power were calibrated to ensure that the RF excitation angle θ was equal to 30° for all observed metabolites in the entire tissue of interest.

2.7 | Data analysis and statistics

^{13}C signal integrals were quantified with Bayes (Washington University, St. Louis, MO), as previously described.³⁰ All metabolite signal integrals were normalized to the respective substrate signal integral. Statistics on the comparison between polarization levels obtained with and without the addition of ethanol to the substrate mixture were computed via 2-tailed Student's t-tests for unpaired data with equal variance. All data are expressed as mean \pm standard error of the mean.

3 | RESULTS

To measure in vivo cardiac metabolism of different substrates simultaneously, without adding persistent radicals, it was demonstrated that copolarization of both ^{13}C -labeled butyric and pyruvic acids is possible using photo-induced nonpersistent radicals. The radicals created at 77 K by UV irradiation of pyruvic acid act as polarizing agents for both acids. The presence of butyric acid did not modify the structure of the UV-induced radical, based on the ESR spectral analysis. Because the radicals are scavenged during the dissolution process, a radical-free hyperpolarized ^{13}C labeled substrate mixture was obtained and could be injected for in vivo real-time metabolic measurements.

To assess the concentration of radicals, ESR measurements were performed at several time points during the irradiation process, which indicated that the maximum concentration of radicals was obtained after 1 hour of irradiation, regardless of the composition of the mixture (Figure 2A). The solution containing solely ^{13}C substrates reached a maximum radical concentration of 16 mM (Figure 2A). The solution in which ethanol was added to a concentration of 20% (v/v) reached a maximum radical concentration of 26 mM (Figure 2A), a significant increase in radical concentration compared with the mixture of only pyruvic and butyric acid.

To assess the degree of achieved hyperpolarization, in vitro hyperpolarized and thermally polarized ^{13}C MRS experiments were performed on both mixtures, and a mean ^{13}C enhancement of 4'100 and 6'400 (Figure 2B) was observed. A ^{13}C mean polarization of $3.3 \pm 0.5\%$ and $5.2 \pm 0.5\%$ (Figure 2C) was measured for the undoped mixture and the ethanol-doped mixture, respectively. The addition of ethanol to the substrate mixture led to a significant increase in the measured polarization level (Figure 2B,C; $P = .03$). As expected, the 2 different methods used to measure the thermally polarized ^{13}C resonances did not lead to significantly different estimations of the ^{13}C polarization (Supporting Information Table S1). Both the HP time courses (Figure 2D) of [$1\text{-}^{13}\text{C}$]pyruvic and [$1\text{-}^{13}\text{C}$]butyric acid as well as the summed spectrum (Figure 2E) show a higher signal intensity of the C1 label of pyruvic acid compared with that of butyric acid, which corresponds to the different concentrations of the compounds when mixed in equal volumes. The difference in enhancement between the thermally polarized substrates versus the hyperpolarized substrates was evident in both spectra (Figure 2E).

To assess whether in vivo metabolism could be observed, the UV-irradiated mixture containing only [$1\text{-}^{13}\text{C}$]pyruvic and [$1\text{-}^{13}\text{C}$]butyric acid was injected in 5 healthy animals. The myocardial metabolism of the hyperpolarized UV-irradiated mixture was followed in real time (Figure 3A),

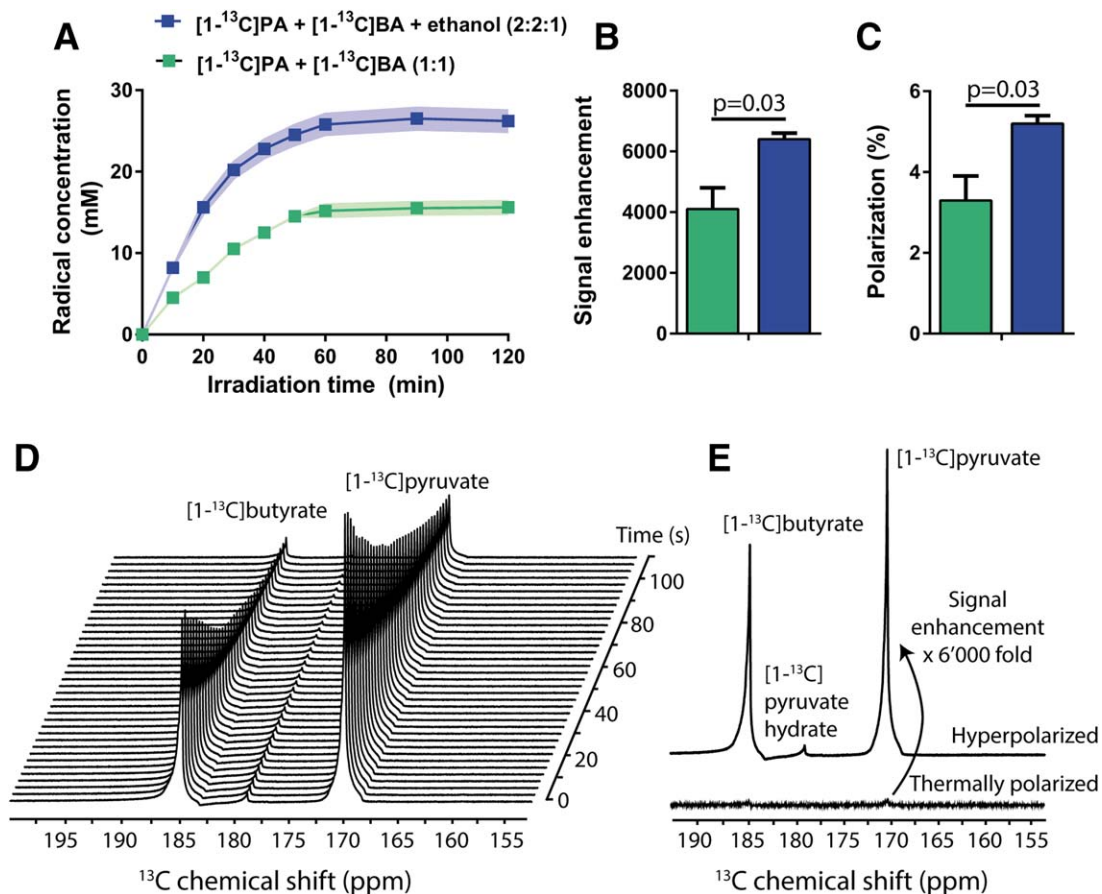


FIGURE 2 A, Radical concentration as a function of the UV light irradiation time for 2 different mixtures: $[1-^{13}\text{C}]\text{pyruvic}$ - and $[1-^{13}\text{C}]\text{butyric}$ acids in a 1:1 volume ratio (green), and the same with the addition of ethanol (blue). Signal intensity enhancement (B) and polarization levels (C) of pyruvic acid using 2 differently prepared mixtures. D, E, In vitro hyperpolarized ^{13}C MRS of the UV-irradiated mixture of ^{13}C labeled substrates. D, Hyperpolarized ^{13}C spectral time course of the mixture measured in the infusion pump inside the MRI scanner after dissolution and automated sample transfer. Hyperpolarized spectra were acquired with an RF excitation angle of 5° and TR of 3 seconds. E, The first measured hyperpolarized ^{13}C spectrum after dissolution (top spectrum) was compared with its thermally polarized counterpart (bottom spectrum) to calculate the SNR enhancement in liquid state. A 6'000-fold signal enhancement was measured. All spectra are displayed without spectral line broadening

showing that sufficient polarization levels were obtained to measure metabolic processes using this radical-free method. The metabolism of hyperpolarized $[1-^{13}\text{C}]\text{pyruvate}$ led to the detection of lactate, alanine, and ^{13}C bicarbonate (Figure 3B). Metabolism of hyperpolarized butyrate resulted in ^{13}C labeling in acetylcarnitine (Figure 3B). The ratio of acetylcarnitine relative to injected substrate butyrate was 0.011 ± 0.004 , the lactate-to-pyruvate ratio was 0.068 ± 0.023 , the alanine-to-pyruvate ratio was 0.029 ± 0.008 , and the bicarbonate-to-pyruvate ratio was 0.006 ± 0.001 (Figure 4).

4 | DISCUSSION

In the present study, we show for the first time the measurement of myocardial metabolism using DNP with photo-induced nonpersistent radicals. Metabolic conversion of the injected substrate mixture resulted in the ^{13}C labeling and in vivo measurement of acetylcarnitine, bicarbonate, and

lactate. Although the ^{13}C polarization levels observed in this study were not as large as those obtained with persistent radicals, we demonstrated that multiple ^{13}C -labeled substrates could be copolarized in UV-irradiated mixtures containing pyruvic and butyric acid, and that the mixture could be used for in vivo cardiac metabolic studies in rats. The use of non-persistent radicals may reduce cost and simplify the preparation process of hyperpolarized ^{13}C -substrate mixtures for in vivo applications, notably by removing the need for a filter, which could also allow reduction of the delay between dissolution and injection. This shows that the noninvasive and simultaneous measurement of metabolic substrate competition can be measured in the heart in a single experiment without the addition of exogenous persistent radicals or glassing solvents, thus facilitating the translation of these types of experiments to humans.³¹

In vivo metabolic measurements using hyperpolarized ^{13}C MRS following the administration of UV-irradiated mixtures were compared with the ^{13}C spectra acquired after

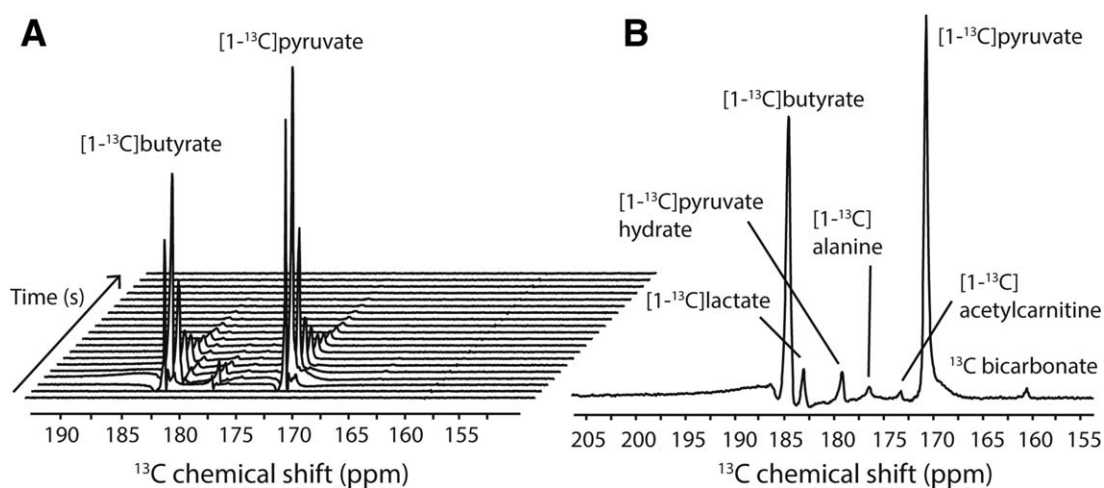


FIGURE 3 In vivo ^{13}C MRS measured in the healthy myocardium after the injection of a hyperpolarized UV-irradiated $[1-^{13}\text{C}]$ pyruvate/ $[1-^{13}\text{C}]$ butyrate mixture. A, Representative ^{13}C spectral time course following the injection of the substrate mixture. Acquisitions were respiratory-gated and cardiac-triggered with a repetition time of 3 seconds and a flip angle of 30° . B, Sum of the 15 spectra acquired starting 9 seconds after dissolution. The myocardial metabolism of both pyruvate and butyrate could be observed through the formation of alanine, lactate, bicarbonate, and acetylcarnitine

injection of mixtures that were hyperpolarized with persistent radicals (TEMPO)¹⁴ to illustrate the effect of the difference in ^{13}C polarization (Figure 5). Although our study with the persistent TEMPO radicals generated a larger ^{13}C polarization ($13 \pm 2\%$ at time of injection¹⁴), similar metabolite ratios were observed in the current study using UV-induced non-persistent radicals, albeit with different SNR levels. Both studies were conducted under identical experimental conditions with in vivo femoral vein injections starting 3 seconds after dissolution. One of the consequences of the larger ^{13}C polarization obtained in our study with the persistent radicals was that the ^{13}C labeling of glutamate and acetoacetate could be observed,¹⁴ but there was no attempt to filter or scavenge the nitroxyl radicals, which were injected together with the substrates. Because of the spectral overlap with metabolites originating from pyruvate metabolism, the resonances of β -hydroxybutyrate and citrate, which have been observed in previous studies using hyperpolarized butyrate and persistent

radicals, could not be observed.^{14,32} However, the wide versatility of photo-induced nonpersistent radicals permits the hyperpolarization of substrate mixtures containing unlabeled pyruvic acid. Because of substrate competition, changes in use may occur with co-infusion of carbohydrates and fatty acids,¹⁴ but this may be exploited to study specific metabolic phenotypes.

The total amount of acetate in the hyperpolarized solution is equivalent to the number of photo-induced radicals (16 mM) created in the frozen sample (100 μL).¹⁶ After dissolution and infusion, we estimate that this increases the blood acetate concentration by 10 μM . Because rats typically have 200 μM acetate in blood,³³ and based on our earlier studies of ^{13}C -labeled acetate conversion to ^{13}C acetylcarnitine in muscle,^{25,26} the minor contribution of unlabeled acetate was considered to have negligible effects on the acetylcarnitine pool size.

The SNR and polarization level obtained in these experiments were sufficient to detect the downstream metabolites acetylcarnitine, bicarbonate and lactate, but a higher SNR would be required for detecting other metabolites and would be beneficial for imaging applications. The liquid-state ^{13}C polarization obtained in the present study was considerably lower ($\sim 3.3\%$) than the maximum achieved previously using TEMPO ($\sim 13\%$).¹⁴ Because nitroxyl radicals show a broader ESR spectrum if compared with the one arising from UV-irradiated pyruvic acid,¹⁷ the lower DNP enhancement must result from the nonoptimal concentration of paramagnetic centers for the temperature and magnetic field strength. Indeed, as illustrated by the addition of ethanol to the pyruvic and butyric acid mixture, increasing the radical concentration from 16 to 26 mM improved the polarization from 3.3 to 5.1%, demonstrating that it is possible to increase the ^{13}C polarization this way. It was already previously reported that diluting pyruvic acid in a polar solvent increased the radical concentration

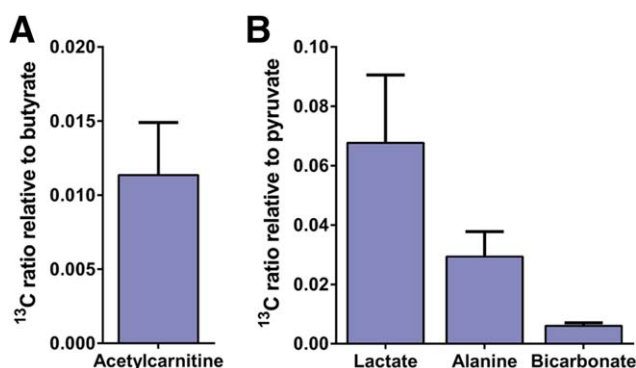


FIGURE 4 ^{13}C metabolite signal integral ratios relative to their respective injected substrate following the injection and myocardial metabolism of a UV-irradiated radical-free mixture of $[1-^{13}\text{C}]$ pyruvic acid and $[1-^{13}\text{C}]$ butyric acid

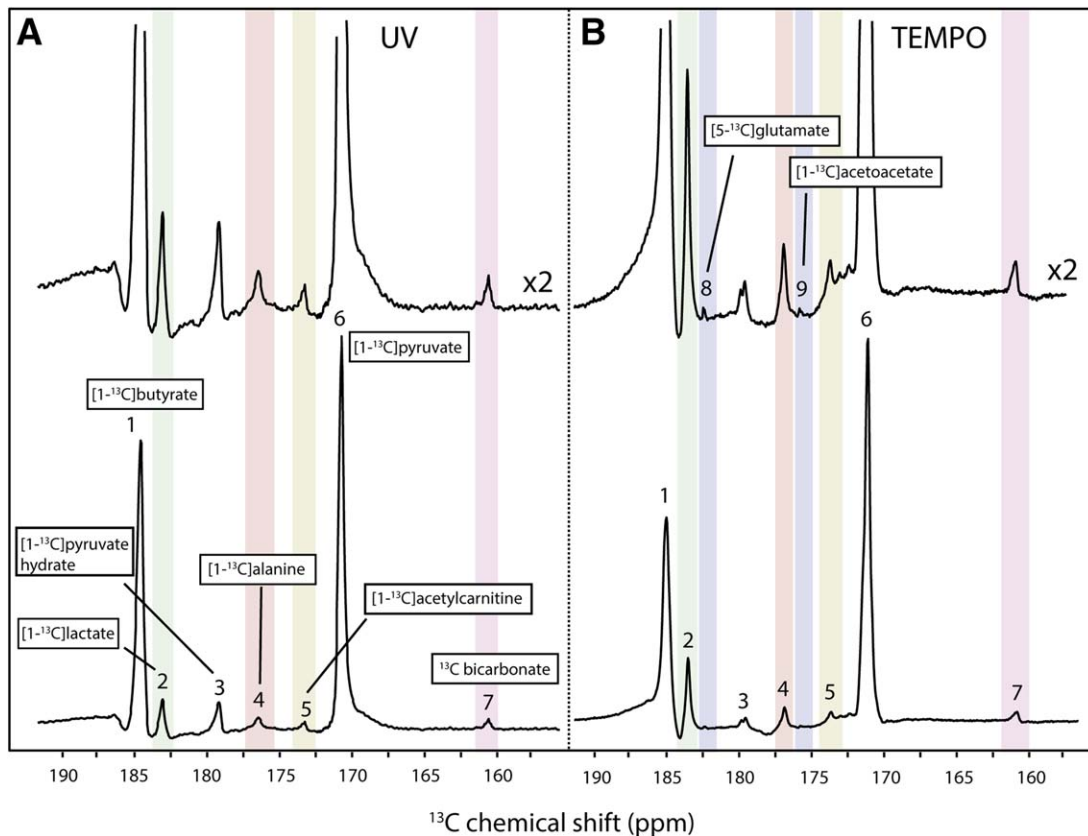


FIGURE 5 Comparison of spectra acquired *in vivo* after the administration of a UV-irradiated mixture containing $[1-^{13}\text{C}]$ pyruvate and $[1-^{13}\text{C}]$ butyrate (left panel) and the administration of a mixture of $[1-^{13}\text{C}]$ pyruvate and $[1-^{13}\text{C}]$ butyrate containing TEMPO persistent radicals (right panel). Data obtained using TEMPO radicals were presented in an earlier publication.¹⁴ Spectra in the top panels were scaled with a factor of 2. The observed metabolites are indicated as follows: 1, $[1-^{13}\text{C}]$ butyrate; 2, $[1-^{13}\text{C}]$ lactate; 3, $[1-^{13}\text{C}]$ pyruvate hydrate; 4, $[1-^{13}\text{C}]$ alanine; 5, $[1-^{13}\text{C}]$ acetylcarnitine; 6, $[1-^{13}\text{C}]$ pyruvate; 7, ^{13}C -bicarbonate; 8, $[5-^{13}\text{C}]$ glutamate; 9, $[1-^{13}\text{C}]$ acetoacetate

above 20 mM.¹⁷ Besides the effect on radical yield, ethanol addition may also enhance polarization by improving the glassing. The increased radical concentration is particularly beneficial to the DNP process when working at high magnetic field, as was done in the current study. However, to avoid any additional metabolic perturbation, the ethanol mixtures were not used for the *in vivo* cardiac experiments.

Preliminary results show that the use of a broadband UV source also dramatically improves the maximum radical yield.¹⁸ At this high field, given the width of the UV-PA radical spectrum compared with TEMPO,¹⁷ modulating the microwave frequency might be beneficial for improving the polarization level and reducing the build-up time.³⁴ However, because all of these potential improvements require additional hardware, the development of these methods to increase the ^{13}C polarization was beyond the scope of this study.

There are several advantages to the use of photo-induced nonpersistent radicals: In addition to cost reduction, the removal of the filtration step would simplify the production process of hyperpolarized substrates for clinical applications and may improve its robustness. In addition, as was recently demonstrated with hyperpolarized $[1-^{13}\text{C}]$ pyruvic acid,¹⁸ the use of photo-induced radicals allows for the storage of

hyperpolarized substrates and may enable their transport to another location, which is of particular interest for clinical studies at sites without DNP equipment. Our results demonstrate the feasibility of preparing transportable copolarized frozen substrate mixtures after annihilation of the nonpersistent radicals in the solid state.

5 | CONCLUSION

We conclude that copolarization of a substrate mixture containing $[1-^{13}\text{C}]$ butyric acid and $[1-^{13}\text{C}]$ pyruvic acid following UV irradiation is possible and leads to a sufficiently high polarization to measure *in vivo* myocardial metabolism. It enables noninvasive and simultaneous monitoring of separate metabolic pathways in a single experiment, without the addition of persistent radicals or glassing solvents.

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CONFLICT OF INTEREST

Arnaud Comment is currently employed by General Electric Medical Systems Inc.

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SUPPORTING INFORMATION

Additional Supporting Information may be found in the supporting information tab for this article.

Table S1 Liquid-state ^{13}C signal enhancements and polarization levels of $[1-^{13}\text{C}]$ pyruvic acid (PA) and $[1-^{13}\text{C}]$ butyric acid (BA) measured inside the separator/infusion pump

Note: Two types of mixtures were subjected to low-temperature UV irradiation: a 1:1 (v/v) mixture of both acids, and a 1:1 (v/v) mixture also containing 20% ethanol by volume. After UV irradiation, the ^{13}C substrate mixtures were hyperpolarized with DNP and automatically transferred to a separator/infusion pump, located inside an MRI scanner, after dissolution. All ^{13}C MRS measurements were performed in liquid state within the separator/infusion pump. Signal enhancements were determined from the maximum measured hyperpolarized signal intensity (peak integral) relative to the thermally polarized signal intensity. Thermal polarization was measured after the complete decay of the hyperpolarized signal with additional nulling performed by the application of RF saturation pulses. Two methods were used to determine the thermally polarized ^{13}C signal intensity as described in the “Methods” section: either by the use of 90° RF excitation pulses with a TR of 240 seconds (Method 1), or the addition of contrast agent and 5° RF excitation pulses with a TR of 1.1 seconds (Method 2). The polarization levels (%) were calculated based on the determined signal enhancements and assuming a thermal equilibrium ^{13}C polarization of 8.1 ppm. No significant differences were found between the two thermal methods ($P = .72$).

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