

# High Magnetic Field Detunes Vibronic Resonances in Photosynthetic Light Harvesting

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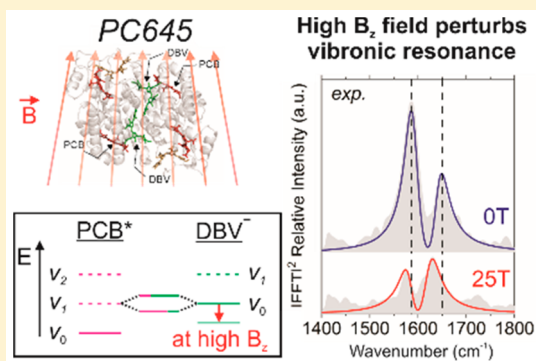
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## Supporting Information

**ABSTRACT:** The origin and role of oscillatory features detected in recent femtosecond spectroscopy experiments of photosynthetic complexes remain elusive. A key hypothesis underneath of these observations relies on electronic–vibrational resonance, where vibrational levels of an acceptor chromophore match the donor–acceptor electronic gap, accelerating the downhill energy transfer. Here we identify and detune such vibronic resonances using a high magnetic field that exclusively shifts molecular exciton states. We implemented ultrafast pump–probe spectroscopy into a specialized 25 T magnetic field facility and studied the light-harvesting complex PC645 from a cryptophyte algae where strongly coupled chromophores form molecular exciton states. We detected a change in high-frequency coherent oscillations when the field was engaged. Quantum chemical calculations coupled with a vibronic model explain the experiment as a magnetic field-induced shift of the exciton states, which in turn affects the electronic–vibrational resonance between pigments within the protein. Our results demonstrate the delicate sensitivity of interpigment coherent oscillations of vibronic origin to electronic–vibrational resonance interactions in light-harvesting complexes.



Reports of coherent beatings in complex systems continue to attract researchers because they suggest a mechanism for tuning photoinduced reactions, e.g., energy transfer, where possible implications have not previously been deeply considered.<sup>1</sup> Coherence therefore has become an engaging topic in recent years.<sup>2–18</sup> Theoretical studies have now predicted that coherent oscillations observed in many ultrafast experiments on light-harvesting complexes<sup>19–23</sup> are assigned to vibronic coherence—quantum mechanical excitation delocalization that depends on the nuclear coordinates of the light-harvesting chromophores.<sup>24–32</sup> This means that the vibrational motion of the chromophores involved becomes correlated by electronic coupling; therefore, the ladders of states of the chromophores do not independently absorb or emit energy. Thus, in many cases, the long-lived coherent oscillations observed in light-harvesting complexes (and charge separation<sup>33</sup>) can be explained by exciton–vibration resonance.

Resonance effects are generally important for photoinduced chemical reactivity because they have pronounced functional consequences. For example, radiationless transitions (e.g.,

internal conversion) are dictated by the energy gap law, where resonant-enhancement of the rate leads to its exponential dependence on the energy gap between the initial and final states.<sup>34</sup> “Flickering resonances” in bridge-mediated electron transfer along DNA strands are predicted to bring dynamically disordered intermediate states into electronic degeneracy, supporting band-like electron transport during a short transient resonance time.<sup>35</sup> The significance of resonance enhancement in photochemistry calls for incisive experimental methods to measure electronic–vibrational resonances and clarify their implication in photosynthetic energy transfer. Recent studies employing two-dimensional electronic spectroscopy (2DES) suggested the presence of electronic–vibrational mixed coherences in various photosynthetic complexes.<sup>33,36,37</sup> However, unequivocal identification of the vibronic coupling and its sensitivity to resonance conditions is a challenging task owing

**Received:** September 6, 2018

**Accepted:** September 10, 2018

**Published:** September 10, 2018

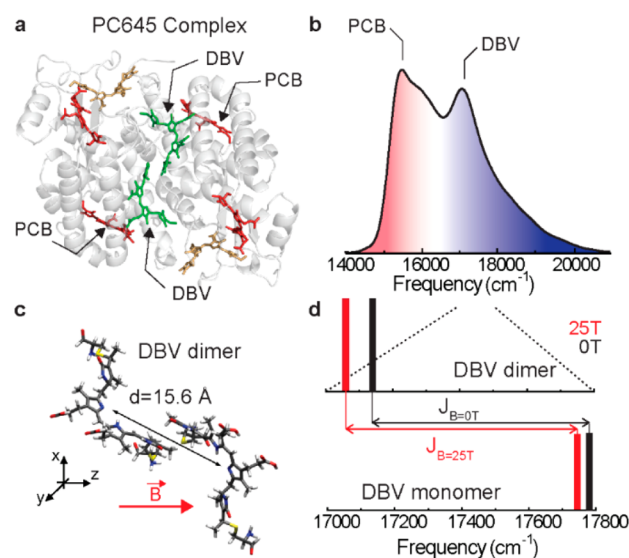
to the complex structure of 2DES signals. A suitable experiment should remove the resonance between the vibrational frequency and the electronic frequency difference between donor and acceptor chromophores and detect the concomitant intensity and energy redistribution initially caused by vibronic mixing. A possible way to remove the electronic resonance could be to exploit protein mutagenesis in such a way as to energetically shift the absorption origin of one chromophore;<sup>38,39</sup> however, a less “invasive” and controllable method is desirable.

Here we demonstrate how a strong external magnetic field can be used to detune vibronic resonance in the light-harvesting complex PC645 isolated from the aquatic cryptophyte algae *Chroomonas mesostigmatica* (CCMP 269). The idea of using an external magnetic field to perturb electronic transitions relies on spin–orbit coupling. An external magnetic field interacts with a molecule through the electronic part of its wave function and has two primary effects on the observed energy levels: a spin splitting of electronic levels (Zeeman splitting) and a diamagnetic shift—an increase in energy of both of the spin-split levels with magnetic field. In a diamagnetic system such as PC645, small static magnetic fields negligibly perturb the molecular absorption spectrum. However, if the magnetic energy becomes comparable to the excitonic coupling (typically at magnetic fields of several Tesla), these fields start to affect the spatial distribution of the orbital electronic wave functions of electrons involved in the photoexcitation. We also assume that the nuclear motion, and hence vibrational spectrum, of a molecule (that is not freely rotating) will not be perturbed by the magnetic field because the magnetic moment generated by the atomic motion is negligible.<sup>40</sup> Thus, a high magnetic field can act as a noninvasive reagent for disentangling electronic–vibrational contributions to interchromophoric interactions.

The *Chroomonas* PC645 light-harvesting complex comprises four chromophore pairs (Figure 1a) that are responsible for the broad absorption in the yellow–orange part of the solar spectrum. These chromophores establish an efficient energy funnel from the core to the periphery of the complex.<sup>41</sup> A prominent pathway involved in the function of this protein is excitation of the highest energy band, at  $\sim 17000\text{ cm}^{-1}$ , of an exciton state where the energy is coherently delocalized across two dihydrobiliverdin (DBV) chromophores in the center of the light-harvesting complex.<sup>42</sup> A direct energy transfer channel from the donor lowest excitonic state ( $\text{DBV}^-$ ) to the peripheral phycocyanobilin (PCB) acceptors (Figure 1b), absorbing below  $\sim 15500\text{ cm}^{-1}$ , dominates, and energy is transferred in about 600 fs.<sup>41,43</sup>

We applied a TDDFT method<sup>44–46</sup> (details in section 1 of the Supporting Information (SI)), to calculate the effect of a strong magnetic field on the excitonic coupling of the PC645 DBV dimer (Figure 1c). We performed a series of calculations to predict the shift in the excitonic coupling at various magnetic field intensities, ranging from 0 to 25 T (Figure S1). The calculated electronic excitation energies of the DBV monomer and dimer at 25 T are shown in Figure 1d and summarized in Table 1. Our calculations of the electronic coupling at zero field agree with previous quantum-chemical calculations.<sup>47</sup>

The predicted spectral shifts induced on the DBV dimer by the magnetic field, however, are too small to be detected by steady-state absorption measurements, given the complexity and disorder within the light-harvesting complex that



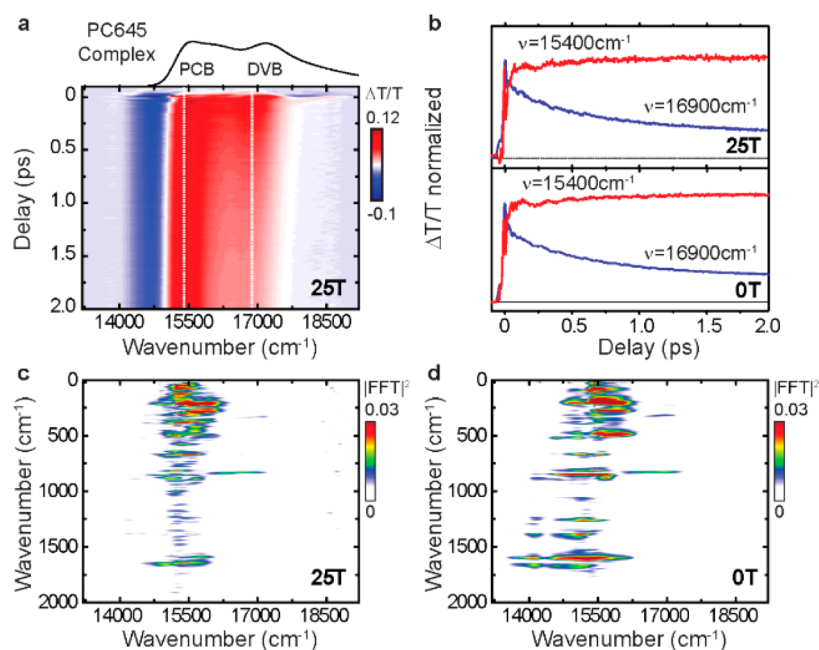
**Figure 1.** (a) Pigment–protein arrangement of the PC645 light-harvesting complex. Four chromophore pairs are identified; of relevance to this study is the central DBV pair (green) and two pairs of PCB chromophores (red). (b) Absorption spectrum of PC645 at 0 T. (c) Molecular structure of the DBV dimer used for TDDFT modeling where the chromophores have a center-to-center separation of 15.6 Å. (d) Calculated excitation energies of the DBV monomer and dimer at a magnetic field  $B_z = 0$  (black) and 25 T (red). These values, shown in Table 1, were used to calculate the shift of the electronic coupling induced by the magnetic field.

**Table 1. Comparison of the Calculated Electronic Excitation Energy ( $E$ ,  $\text{cm}^{-1}$ ) and Electronic Coupling  $J$  of the DBV Monomer and Dimer at  $B_z = 0$  and 25 T**

	$E$ ( $B_z = 0\text{ T}$ ) ( $\text{cm}^{-1}$ )	$E$ ( $B_z = 25\text{ T}$ ) ( $\text{cm}^{-1}$ )
monomer	17780	17744
dimer	17140	17060
$J$	640	684
$\Delta J$		$\sim 44$

contribute to the broad line shapes observed. Instead, we exploit femtosecond broad-band pump–probe spectroscopy, which offers excellent sensitivity to quantum state splittings (vibrational or electronic) detected in the time domain and conveniently brought into the frequency domain by Fourier transformation.

We utilized the Florida Split-Helix magnet<sup>48</sup> developed at the National High Magnetic Field Laboratory (NHMFL) facility for generating the high magnetic fields used in these experiments. The Split-Helix is one of the largest magnets in the world, specifically designed for advanced optical experiments at magnetic fields up to 25 T. Our combined broad-band pump–probe/Split-Helix system offers the unique capability to couple strong static magnetic fields with sub-20 fs laser pulses (details of the experimental apparatus are given in section 2 of the SI). Coherence-sensitive experiments in such high magnetic fields are particularly challenging owing to the sometimes weak oscillatory signals and multiple sources of noise associated with generating such high magnetic fields (i.e., the temperature variation and the high water pressure needed to cool down the magnet during the experiment), which affects the signal-to-noise ratio of our measurements. Nevertheless, a test experiment at 25 T on a reference dye, cresyl violet, proves that high-sensitivity measurements are feasible. The broad-



**Figure 2.** (a) Pump–probe  $\Delta T/T$  map of PC645 measured at 25 T and corresponding region of the linear absorption. (b) Selected pump–probe traces at DBV ( $16900\text{ cm}^{-1}$ ) and PCB ( $15400\text{ cm}^{-1}$ ) probe energies at 0 and 25 T. Fourier transform maps at (c) 25 and (d) 0 T.

band laser pump stimulates vibrational wavepackets that are observed as oscillations superimposed on the pump–probe dynamics. By Fourier transforming the residual oscillations following subtraction of the data with a biexponential fitting function, we retrieve the frequency response of the system. The strongest vibrational mode of cresyl violet has a frequency of  $590\text{ cm}^{-1}$ <sup>49</sup> and is unaffected by the 25 T field, supporting our hypothesis that strong magnetic fields do not alter nuclear motions (Figure S4).

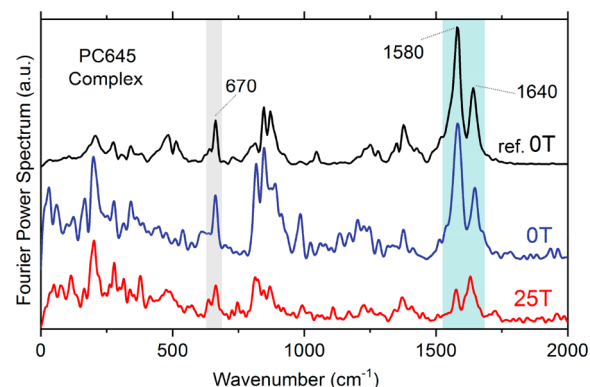
To further validate our hypothesis, we also report a control study on a cryptophyte algae protein subunit containing a single PCB chromophore attached to the  $\alpha$ -subunit, which similarly shows no changes in the ultrafast coherent response when the 25 T magnetic field is engaged (Figure S5). The control experiment on the subunit (Figure S5) reveals two intramolecular vibrations at  $670$  and  $1630\text{ cm}^{-1}$ . Details on the PC645 and  $\alpha$ -subunit sample preparation are described in section 10 of the SI.

Figure 2a shows the pump–probe differential transmission ( $\Delta T/T$ ) map recorded for PC645 in solution at 25 T and ambient temperature. A similar measurement at 0 T is reported in the SI (Figure S6). The signal in both cases is dominated by instantaneous ground-state bleaching of the optically excited transitions of different chromophores in the protein. The  $16800$ – $17800\text{ cm}^{-1}$  probe range displays a subpicosecond decaying component, assigned to ultrafast energy transfer from the DVB donor state to the PCB acceptor excited states.

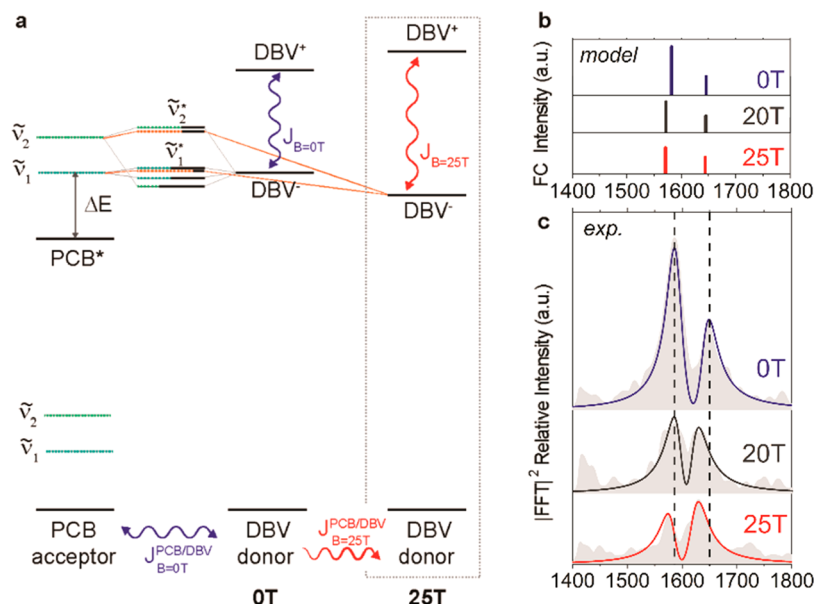
Figure 2b shows the decay of the DBV signature ( $16900\text{ cm}^{-1}$ ) and the corresponding growth of the PCB bleach signal ( $15400\text{ cm}^{-1}$ ) at both 0 and 25 T, which we assigned to ultrafast energy transfer from the DVB donor state to the PCB acceptor excited states. We find that, even though the vibronic coupling effects that we detected previously using 2D electronic spectroscopy<sup>43</sup> are perturbed by the magnetic field (vide infra), there is no magnetic field dependence on the DBV to PCB energy transfer rate. These observations together

support the theoretical study reported by Blau and co-workers.<sup>50</sup>

We applied the same procedure described for the cresyl violet dye to obtain the Fourier-transformed power maps at 25 and 0 T (Figure 2c,d). The clear pattern of coherences spanning from  $200$  to  $1700\text{ cm}^{-1}$ , centered predominantly at the PCB region of the probe spectrum, is preserved when the magnetic field is engaged. In order to analyze the coherent signals at zero field and 25 T, we integrated the power maps in Figure 2c,d over all probe wavelengths. The results are reported in Figure 3, where the 0 and 25 T power spectra measured at the NHMFL (blue and red curves, respectively) are displaced together with a reference spectrum (labeled as



**Figure 3.** Integrated Fourier power spectra of PC645 at 25 and 0 T (red and blue curves, respectively) obtained at the NHMFL and compared with the integrated Fourier power spectrum from a reference experiment at 0 T performed on the PC645 complex under the same experimental conditions at Princeton University laboratories. The shaded regions show signatures of the Fourier spectra, which we focus on in this work, specifically, the lack of modulation of the  $670\text{ cm}^{-1}$  vibrational mode, as well as the marked changes in the high-frequency region between  $1500$  and  $1700\text{ cm}^{-1}$ .



**Figure 4.** (a) Energy level schemes of the vibronically coupled DBV/PCB chromophores. Solid black lines represent electronic states, while the dotted green lines represent the vibrational states of the PCB; vibronic states are labeled as  $\nu^*$ .  $J_B$  is the electronic coupling between the two DBV excitonic states;  $J_{PCB/DBV}$  is the electronic coupling between the DBV and the PCB chromophore, and it weakly depends on the magnetic field. (b) Simulated amplitudes of bands associated with the two oscillations  $\nu_1^*$  and  $\nu_2^*$  at 0, 20, and 25 T, obtained from summation of the amplitudes associated with all Feynman pathways contributing to each coherence<sup>43</sup> (details are discussed in section 11 in the SI). (c) Expansion of measured integrated Fourier power spectra (gray shadows) in the high-frequency spectral region at 0, 20, and 25 T. Solid lines are obtained by fitting the filtered oscillations with the sum of two damped cosines.

“ref.”, black curve in Figure 3) obtained from a similar pump–probe experiment on PC645 in a standard ultrafast laboratory.

At 0 T, strong coherent oscillations in the 200–350 cm<sup>-1</sup> region as well as distinct frequencies at around 490, 670, 850, 1580, and 1640 cm<sup>-1</sup> are evident. While there are some clear differences in the 0 T experiment on PC645 at the NHMFL and the reference measurement, particularly at 800 and 1000 cm<sup>-1</sup> and in the low-frequency region, we exclude changes in these frequency regions on the basis of setup-to-setup reproducibility and instead focus on the power spectrum signatures that reproduce well the reference experiment, namely, those at 670 cm<sup>-1</sup> and between 1500 and 1700 cm<sup>-1</sup>. At these oscillation frequencies, the 0 T experiment on PC645 at the NHMFL matches very well the reference measurement (blue and black curves in Figure 4), confirming that the sensitivity at the NHMFL facility is adequate. The 25 T data (red curve in Figure 4) are largely consistent with the zero-field experiments; as expected, the spectral positions of some of the largest peaks in the power spectrum are primarily unperturbed by the applied field, i.e., the position of the 670 cm<sup>-1</sup> peak is preserved. However, a striking and reproducible change in the Fourier spectra intensity at around 1580–1600 cm<sup>-1</sup> is detected. Specifically, the amplitudes and locations of the 1580 and 1640 cm<sup>-1</sup> bands are somewhat sensitive to the magnetic field. Interestingly, the 1580 cm<sup>-1</sup> frequency, previously assigned to a C=N stretching mode of the PCB chromophore,<sup>51–53</sup> appears only as a small shoulder in the  $\alpha$ -subunit data (Figure S4).

We rationalize the amplitude difference of the 1580 cm<sup>-1</sup> oscillation in the protein as a consequence of an amplification mechanism that becomes active when the chromophores interact closely due to their spatial arrangement in the protein scaffold. This hypothesis has been recently described on the basis of 2DES studies on the PC645 complex.<sup>43</sup> It was

proposed that the vibrational coherence experiences a coherent amplification through vibronic coupling between the energetically remote DBV and PCB chromophores, caused by resonance of the intramolecular PCB vibration with the electronic energy gap between the lower excitonic DBV<sup>-</sup> state and the excited PCB state.<sup>43</sup> The response of these oscillations to the magnetic field suggests that the shift of the excitonic DBV<sup>-</sup> band could be affecting those coherences via changes in vibronic coupling to PCB.

We use a simple vibronic coupling model, like that used previously to analyze 2D electronic spectra recorded for PC645,<sup>43</sup> to qualitatively predict how the coherence amplitudes of vibrations of the PCB chromophores are perturbed by mixing with the electronic DBV<sup>-</sup> state (for details, see section 7 in the SI). In our model (Figure 4a), we consider the DBV<sup>-</sup> exciton state, the PCB excited state, and two local PCB vibrations, at  $\nu_1 = 1560$  cm<sup>-1</sup> and  $\nu_2 = 1640$  cm<sup>-1</sup>. We do not include the DBV vibrations in the model as those vibrationally excited states are not close enough in energy to the PCB acceptor vibrational ground state for appreciable Franck–Condon overlap.

The coupling between these quasi-resonant DBV excitons and PCB vibrationally excited  $S_1$  leads to two vibronic eigenstates ( $\nu_1^*$  and  $\nu_2^*$ ) in the relevant energy window. The degree of vibronic mixing depends on (i) the detuning between the vibrational frequency and the electronic energy gap  $\Delta E$  (fixed at 1560 cm<sup>-1</sup> in resonance with  $\nu_1$ ), (ii) the Huang–Rhys factor of the vibrational modes ( $S = 0.08$ ), and (iii) the electronic coupling between the DBV exciton state and PCB ( $J_{PCB/DBV} = 44$  cm<sup>-1</sup> at 0 T and 48 cm<sup>-1</sup> at 25 T, as calculated in section 6 in the SI).

In order to compare our experimental trend with simulations based on our hypothesized model, Figure 4b (vide infra), we fit the frequency-domain-filtered oscillations, recorded at the

various magnetic field strengths, in the time domain with a sum of two damped cosines. The fitting results are summarized in Table S2 in the SI. The relative amplitudes of the 1580 versus 1640  $\text{cm}^{-1}$  oscillations are clearly magnetically field-dependent, Figure 4c.

At zero field, we found that vibronic mixing of the  $\text{DBV}^-$  with the two PCBs yields states with strong intensity at 1580  $\text{cm}^{-1}$  and a less intense band at 1645  $\text{cm}^{-1}$  (black bars in Figure 4b). When we simulated the effect of the 25 T field tuning the excitonic  $\text{DBV}^-$  state away from resonance (changing  $J_B$  by  $\sim 44 \text{ cm}^{-1}$ ), we found a significant change in the lower vibronic eigenstate. Both vibronic bands undergo a small red shift (by  $\sim 10 \text{ cm}^{-1}$ , red bars in Figure 4b), and the intensity of the 1580  $\text{cm}^{-1}$  band measured at 0 T is strongly decreased at 25 T. The amplitude of the 1640  $\text{cm}^{-1}$  is much less sensitive to the excitonic detuning because of the larger energy detuning  $\Delta E$  compared to the vibrational frequency  $\nu_2$ , although its slight red shift is captured well by our model. This interpretation is supported by the pump–probe data recorded at an intermediate magnetic field of 20 T (Figure S6) and further highlights the favorable resonance conditions of the 1580  $\text{cm}^{-1}$  vibrational mode at ambient conditions. Our calculations are in reasonable agreement with the trend evident in experimental results reported in Figure 4c, which shows an expanded view of the experimental power spectra described in Figure 4 for 0 and 25 T, together with the results at 20 T.

We have found that a very strong magnetic field can perturb the excitonic electronic–vibrational resonances of electronically coupled chromophores in diamagnetic systems. For the first time, we have combined an intense static high magnetic field with ultrafast broad-band spectroscopy to inspect vibronic coherences in a photosynthetic complex. We find that the vibronic tuning markedly affects the amplitude of the 1580  $\text{cm}^{-1}$  band. We found that detuning at high field strengths of this vibronic resonance does not affect the energy transfer rate, as shown in Figure S8. That is, even though the vibronic coupling effects that we detected previously using 2D electronic spectroscopy<sup>43</sup> are perturbed by the magnetic field (vide infra), there is no magnetic field dependence on the  $\text{DBV}$  to PCB energy transfer rate. These observations together support the theoretical study reported by Blau and co-workers<sup>50</sup> that find that the spectral line broadening of PC645 is too large relative to the electronic couplings for the vibronic coupling to affect the energy transfer rate. Nevertheless, this unique magnetic field effect helps reveal the complex network of couplings responsible for not only driving energy transfer among pigments within the protein but also tuning the observed frequencies and intensities of coherences in the experiment. These are built first from excitonic coupling of the  $\text{DBV}$  donor pair, which subsequently affects the downstream vibronic coupling with the PCB acceptors.

Our work reveals the sensitivity of vibronic coupling to electronic–vibrational resonance as the field shifts the electronic donor state relative to the acceptor vibrational states. This result indicates that controlled chromophore–chromophore interactions for light harvesting may involve a surprisingly precise balance of energy gaps. This approach may contribute to framing the mechanisms of vibronically assisted photoinduced dynamics and use them to illuminate design principles for directed light harvesting.

## ■ ASSOCIATED CONTENT

### 📄 Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.jpcllett.8b02748.

Real-time TDDFT, detailed discussion of the experimental design, various control broad-band pump–probe experiments at zero and high magnetic fields, coupling calculations for the vibronic coupling model, vibronic coupling calculations, Fourier analysis and calculation of coherence amplitudes for PC645, population dynamics at 25 T, and the preparation method for the PC645 protein and  $\alpha$ -subunit (PDF)

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### Notes

The authors declare no competing financial interest.

## ■ ACKNOWLEDGMENTS

The authors acknowledge financial support by the National Science Foundation, MRI No. DMR-1229217, and by Princeton University through the Innovation Fund for New Ideas in the Natural Sciences. A portion of this work was performed at the National High Magnetic Field Laboratory, which is supported by the National Science Foundation Cooperative Agreement No. DMR-1157490 and the State of Florida. M.M. acknowledges financial support by European Community (H2020 Marie Skłodowska-Curie Actions), Project No. 655059. M.B.O. acknowledges financial support by the National Science Foundation - Institute for Complex Adaptive Matter (NSF-ICAM) and the National Science Foundation for the use of supercomputing resources through the Extreme Science and Engineering Discovery Environment (XSEDE), Project No. TG-CHE140097. B.K. acknowledges support by the National Science Foundation Graduate Research Fellowship under Grant Number DGE-1656466. M.M., J.C.D., M.B., and S.A.G. acknowledge Dmitry Semenov for valuable support during the magnet time.

## ■ REFERENCES

- (1) Chenu, A.; Scholes, G. D. Coherence in Energy Transfer and Photosynthesis. *Annu. Rev. Phys. Chem.* **2015**, *66*, 69–96.
- (2) Scholes, G. D.; Fleming, G. R.; Chen, L. X.; Aspuru-Guzik, A.; Buchleitner, A.; Coker, D. F.; Engel, G. S.; van Grondelle, R.; Ishizaki, A.; Jonas, D. M.; et al. Using Coherence to Enhance Function in Chemical and Biophysical Systems. *Nature* **2017**, *543*, 647–656.
- (3) Romero, E.; Novoderezhkin, V. I.; van Grondelle, R. Quantum Design of Photosynthesis for Bio-inspired Solar-energy Conversion. *Nature* **2017**, *543*, 355–365.
- (4) Caprasecca, S.; Guido, C. A.; Mennucci, B. Control of Coherences and Optical Responses of Pigment–Protein Complexes by Plasmonic Nanoantennae. *J. Phys. Chem. Lett.* **2016**, *7*, 2189–2196.
- (5) Rury, A. S.; Sorenson, S. A.; Dawlaty, J. M. Evidence of Ultrafast Charge Transfer Driven by Coherent Lattice Vibrations. *J. Phys. Chem. Lett.* **2017**, *8*, 181–187.

- (6) Perlík, V.; Lincoln, C.; Šanda, F.; Hauer, J. Distinguishing Electronic and Vibronic Coherence in 2D Spectra by Their Temperature Dependence. *J. Phys. Chem. Lett.* **2014**, *5*, 404–407.
- (7) Ghosh, S.; Bishop, M. M.; Roscioli, J. D.; LaFountain, A. M.; Frank, H. A.; Beck, W. F. Excitation Energy Transfer by Coherent and Incoherent Mechanisms in the Peridinin–Chlorophyll *a* Protein. *J. Phys. Chem. Lett.* **2017**, *8*, 463–469.
- (8) Duan, H.-G.; Miller, R. J. D.; Thorwart, M. Impact of Vibrational Coherence on the Quantum Yield at a Conical Intersection. *J. Phys. Chem. Lett.* **2016**, *7*, 3491–3496.
- (9) Mallus, M. I.; Aghtar, M.; Chandrasekaran, S.; Lüdemann, G.; Elstner, M.; Kleinekathöfer, U. Relation between Dephasing Time and Energy Gap Fluctuations in Biomolecular Systems. *J. Phys. Chem. Lett.* **2016**, *7*, 1102–1108.
- (10) Dutta, R.; Bagchi, B. Environment-Assisted Quantum Coherence in Photosynthetic Complex. *J. Phys. Chem. Lett.* **2017**, *8*, 5566–5572.
- (11) Morrison, A. F.; Herbert, J. M. Evidence for Singlet Fission Driven by Vibronic Coherence in Crystalline Tetracene. *J. Phys. Chem. Lett.* **2017**, *8*, 1442–1448.
- (12) Ma, F.; Romero, E.; Jones, M. R.; Novoderezhkin, V. I.; van Grondelle, R. Vibronic Coherence in the Charge Separation Process of the *Rhodobacter sphaeroides* Reaction Center. *J. Phys. Chem. Lett.* **2018**, *9*, 1827–1832.
- (13) Ma, F.; Yu, L.-J.; Hendriks, R.; Wang-Otomo, Z.-Y.; van Grondelle, R. Excitonic and Vibrational Coherence in the Excitation Relaxation Process of Two LH1 Complexes as Revealed by Two-Dimensional Electronic Spectroscopy. *J. Phys. Chem. Lett.* **2017**, *8*, 2751–2756.
- (14) Izmaylov, A. F.; Joubert-Doriol, L. Quantum Nonadiabatic Cloning of Entangled Coherent States. *J. Phys. Chem. Lett.* **2017**, *8*, 1793–1797.
- (15) Groenhof, G.; Toppari, J. J. Coherent Light Harvesting through Strong Coupling to Confined Light. *J. Phys. Chem. Lett.* **2018**, *9*, 4848–4851.
- (16) Wang, H.; Valkunas, L.; Cao, T.; Whittaker-Brooks, L.; Fleming, G. R. Coulomb Screening and Coherent Phonon in Methylammonium Lead Iodide Perovskites. *J. Phys. Chem. Lett.* **2016**, *7*, 3284–3289.
- (17) Monahan, D. M.; Guo, L.; Lin, J.; Dou, L.; Yang, P.; Fleming, G. R. Room-Temperature Coherent Optical Phonon in 2D Electronic Spectra of CH<sub>3</sub>NH<sub>3</sub>PbI<sub>3</sub> Perovskite as a Possible Cooling Bottleneck. *J. Phys. Chem. Lett.* **2017**, *8*, 3211–3215.
- (18) Fujita, T.; Atahan-Evrenk, S.; Sawaya, N. P. D.; Aspuru-Guzik, A. Coherent Dynamics of Mixed Frenkel and Charge-Transfer Excitons in Dinaphtho[2,3-b:2'3'-f]thieno[3,2-b]-thiophene Thin Films: The Importance of Hole Delocalization. *J. Phys. Chem. Lett.* **2016**, *7*, 1374–1380.
- (19) Engel, G. S.; Calhoun, T. R.; Read, E. L.; Ahn, T. K.; Mančal, T.; Cheng, Y.-C.; Blankenship, R. E.; Fleming, G. R. Evidence for Wavelike Energy Transfer through Quantum Coherence in Photosynthetic Systems. *Nature* **2007**, *446*, 782–786.
- (20) Collini, E.; Wong, C. Y.; Wilk, K. E.; Curmi, P. M. G.; Brumer, P.; Scholes, G. D. Coherently Wired Light-harvesting in Photosynthetic Marine Algae at Ambient Temperature. *Nature* **2010**, *463*, 644–647.
- (21) Panitchayangkoon; Hayes, G.; Fransted, K. A.; Caram, J. R.; Harel, E.; Wen, J.; Blankenship, R. E.; Engel, G. S. Long-lived Quantum Coherence in Photosynthetic Complexes at Physiological Temperature. *Proc. Natl. Acad. Sci. U. S. A.* **2010**, *107*, 12766–12770.
- (22) Mirkovic, T.; Ostroumov, E. E.; Anna, J. M.; van Grondelle, R.; Govindjee; Scholes, G. D. Light Absorption and Energy Transfer in the Antenna Complexes of Photosynthetic Organisms. *Chem. Rev.* **2017**, *117*, 249–293.
- (23) Christensson, N.; Kauffmann, H. F.; Pullerits, T.; Mancal, T. Origin of Long-Lived Coherences in Light-harvesting Complexes. *J. Phys. Chem. B* **2012**, *116*, 7449–7454.
- (24) Kolli, A.; O'Reilly, E. J.; Scholes, G. D.; Olaya-Castro, A. The Fundamental Role of Quantized Vibrations in Coherent Light Harvesting by Cryptophyte Algae. *J. Chem. Phys.* **2012**, *137*, 174109.
- (25) Chenu, A.; Christensson, N.; Kauffmann, H. F.; Mancal, T. Enhancement of Vibronic and Ground-state Vibrational Coherences in 2D Spectra of Photosynthetic Complexes. *Sci. Rep.* **2013**, *3*, 2029.
- (26) Tiwari, V.; Peters, W. K.; Jonas, D. M. Electronic Resonance with Anticorrelated Pigment Vibrations Drives Photosynthetic Energy Transfer Outside the Adiabatic Framework. *Proc. Natl. Acad. Sci. U. S. A.* **2013**, *110*, 1203–1208.
- (27) Chin, A. W.; Prior, J.; Rosenbach, R.; Caycedo-Soler, F.; Huelga, S. F.; Plenio, M. B. The Role of Non-equilibrium Vibrational Structures in Electronic Coherence and Recoherence in Pigment-Protein Complexes. *Nat. Phys.* **2013**, *9*, 113–118.
- (28) Plenio, M. B.; Almeida, J.; Huelga, S. F. Origin of Long-lived Oscillations in 2D-spectra of a Quantum Vibronic Model: Electronic Versus Vibrational Coherence. *J. Chem. Phys.* **2013**, *139*, 235102.
- (29) Lim, J.; Paleček, D.; Caycedo-Soler, F.; Lincoln, C. N.; Prior, J.; von Berlepsch, H.; Huelga, S. F.; Plenio, M. B.; Zigmantas, D.; Hauer, J. Vibronic Origin of Long-lived Coherence in an Artificial Molecular Light Harvester. *Nat. Commun.* **2015**, *6*, 7755.
- (30) Chin, A. W.; Datta, A.; Caruso, F.; Huelga, S. F.; Plenio, M. B. Noise-assisted Energy Transfer in Quantum Networks and Light-harvesting Complexes. *New J. Phys.* **2010**, *12*, 065002.
- (31) Womick, J. M.; Moran, A. M. Vibronic Enhancement of Exciton Sizes and Energy Transport in Photosynthetic Complexes. *J. Phys. Chem. B* **2011**, *115*, 1347–1356.
- (32) Prior, J.; Chin, A. W.; Huelga, S. F.; Plenio, M. B. Efficient Simulation of Strong System-Environment Interactions. *Phys. Rev. Lett.* **2010**, *105*, 050404.
- (33) Romero, E.; Augulis, R.; Novoderezhkin, V. I.; Ferretti, M.; Thieme, J.; Zigmantas, D.; van Grondelle, R. Quantum Coherence in Photosynthesis for Efficient Solar-Energy Conversion. *Nat. Phys.* **2014**, *10*, 676–683.
- (34) Freed, K. F. *Radiationless Processes in Molecules and Condensed Phases: in Molecules and Condensed Phases*; Springer: Berlin, Heidelberg, 1976; pp 23–168.
- (35) Zhang, Y.; Liu, C.; Balaeff, A.; Skourtis, S. S.; Beratan, D. N. Biological Charge Transfer via Flickering Resonance. *Proc. Natl. Acad. Sci. U. S. A.* **2014**, *111*, 10049–10054.
- (36) Womick, J. M.; West, B. A.; Scherer, N. F.; Moran, A. M. Vibronic Effects in the Spectroscopy and Dynamics of C-phycocyanin. *J. Phys. B: At., Mol. Opt. Phys.* **2012**, *45*, 154016.
- (37) Fuller, F. D.; Pan, J.; Gelzinis, A.; Butkus, V.; Senlik, S. S.; Wilcox, D. E.; Yocum, C. F.; Valkunas, L.; Abramavicius, D.; Ogilvie, J. P. Vibronic Coherence in Oxygenic Photosynthesis. *Nat. Chem.* **2014**, *6*, 706–711.
- (38) Fowler, G. J. S.; Visschers, R. W.; Grief, G. G.; van Grondelle, R.; Hunter, C. N. Genetically Modified Photosynthetic Antenna Complexes with Blueshifted Absorbency Bands. *Nature* **1992**, *355*, 848–850.
- (39) Maiuri, M.; Ostroumov, E. E.; Saer, R. G.; Blankenship, R. E.; Scholes, G. D. Coherent wavepackets in the Fenna-Matthews-Olson complex are robust to excitonic-structure perturbations caused by mutagenesis. *Nat. Chem.* **2018**, *10*, 177–183.
- (40) Canters, G. W.; Van der Waals, J. H. *The Porphyrins*; Academic Press: New York, 1978; pp 531–582.
- (41) Marin, A.; Doust, A. B.; Scholes, G. D.; Wilk, K. E.; Curmi, P. M. G.; van Stokkum, I. H. M.; van Grondelle, R. Flow of Excitation Energy in the Cryptophyte Light-Harvesting Antenna Phycocyanin 645. *Biophys. J.* **2011**, *101*, 1004–1013.
- (42) Mirkovic, T.; Doust, A. B.; Kim, J.; Wilk, K. E.; Curutchet, C.; Mennucci, B.; Cammi, R.; Curmi, P. M. G.; Scholes, G. D. Ultrafast Light Harvesting Dynamics in the Cryptophyte Phycocyanin 645. *Photochem. Photobiol. Sci.* **2007**, *6*, 964–975.
- (43) Dean, J. C.; Mirkovic, T.; Toa, Z. S. D.; Oblinsky, D. G.; Scholes, G. D. Vibronic Enhancement of Algae Light Harvesting. *Chem.* **2016**, *1*, 858–872.

(44) Andrade, X.; Strubbe, D.; De Giovannini, U.; Larsen, A. H.; Oliveira, M. J.; Alberdi-Rodriguez, J.; Varas, A.; Theophilou, I.; Helbig, N.; Verstraete, M. J.; et al. Real-space Grids and the Octopus Code as Tools for the Development of New Simulation Approaches for Electronic Systems. *Phys. Chem. Chem. Phys.* **2015**, *17*, 31371–31396.

(45) Marques, M. A. L.; Castro, A.; Bertsch, G. F.; Rubio, A. Octopus: A First-Principles Tool for Excited Electron–Ion Dynamics. *Comput. Phys. Commun.* **2003**, *151*, 60–78.

(46) Castro, A.; Appel, H.; Oliveira, M.; Rozzi, C. A.; Andrade, X.; Lorenzen, F.; Marques, M. A. L.; Gross, E. K. U.; Rubio, A. Octopus: A Tool for the Application of Time-dependent Density Functional Theory. *Phys. Status Solidi B* **2006**, *243*, 2465–2488.

(47) Harrop, S. J.; Wilk, K. E.; Dinshaw, R.; Collini, E.; Mirkovic, T.; Teng, C. Y.; Oblinsky, D. G.; Green, B. R.; Hoef-Emden, K.; Hiller, R. G.; et al. Single-residue Insertion Switches the Quaternary Structure and Exciton States of Cryptophyte Light-harvesting Proteins. *Proc. Natl. Acad. Sci. U. S. A.* **2014**, *111*, E2666–2675.

(48) Toth, J.; Bird, M. D. FEA-Aided Design for a Working Model of a Split Florida-Helix. *IEEE Trans. Appl. Supercond.* **2008**, *18*, 575–578.

(49) Fuji, T.; Saito, T.; Kobayashi, T. Dynamical Observation of Duschinsky Rotation by Sub-5-fs Real-time Spectroscopy. *Chem. Phys. Lett.* **2000**, *332*, 324–330.

(50) Blau, S. M.; Bennett, D. I. G.; Kreisbeck, C.; Scholes, G. D.; Aspuru-Guzik, A. Local Protein Solvation Drives Direct Down-conversion in Phycobiliprotein PC645 via Incoherent Vibronic Transport. *Proc. Natl. Acad. Sci. U. S. A.* **2018**, *115*, E3342–E3350.

(51) Arpin, P. C.; Turner, D. B.; McClure, S. D.; Jumper, C. C.; Mirkovic, T.; Challa, J. R.; Lee, J.; Teng, C. Y.; Green, B. R.; Wilk, K. E.; et al. Spectroscopic Studies of Cryptophyte Light Harvesting Proteins: Vibrations and Coherent Oscillations. *J. Phys. Chem. B* **2015**, *119*, 10025–10034.

(52) Kneip, C.; Parbel, A.; Foerstendorf, H.; Scheer, H.; Siebert, F.; Hildebrandt, P. Fourier Transform Near-infrared Resonance Raman Spectroscopic Study of the Alpha-subunit of Phycoerythrocyanin and Phycocyanin from the Cyanobacterium *Mastigocladus Laminosus*. *J. Raman Spectrosc.* **1998**, *29*, 939–944.

(53) Andel, F.; Murphy, J. T.; Haas, J. A.; McDowell, M. T.; van der Hoef, I.; Lugtenburg, J.; Lagarias, J. C.; Mathies, R. A. Probing the Photoreaction Mechanism of Phytochrome through Analysis of Resonance Raman Vibrational Spectra of Recombinant Analogues. *Biochemistry* **2000**, *39*, 2667–2676.