

Synthesis and Characterization of Xylazine Hydrochloride Polymorphs, Hydrates, and Cocrystals: A ³⁵Cl Solid-State NMR and DFT Study

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ABSTRACT: Xv	lazine HCl (X) is a veterinarv	analges	sic with many known solid	

forms, making it an ideal system for studying the noncovalent interactions, such as hydrogen bonding, that provide stability to polymorphs, solvates/hydrates, and cocrystal of pharmaceuticals. Herein, we report methods for the reliable preparation and interconversion of polymorphs of X (including mechanochemical pathways), the discovery of a novel polymorph, and the synthesis of three cocrystals with coformers containing amide and carboxylic acid moieties. An understanding of ball milling protocols is essential for optimizing these reactions and ensuring clean and reproducible syntheses of the products in high yields. All materials were characterized using thermal analysis, powder and single-crystal X-ray diffraction (PXRD and SCXRD), and multinuclear solid-state NMR (SSNMR) spectroscopy. ³⁵Cl SSNMR is highlighted for its versatility for fingerprinting polymorphs, hydrates, and cocrystals (including the detection of impurity phases that are not



always evident from PXRD and offering an avenue for optimizing synthetic protocols) and providing molecular-level structural information. The ³⁵Cl electric field gradient (EFG) tensor is extremely sensitive to the unique hydrogen-bonding network in each solid form of X, resulting in distinct powder patterns. Dispersion-corrected plane-wave density functional theory (DFT) structural refinements yield better models of the hydrogen-bonding environments of the chloride ions than is possible through XRD methods alone. Calculations employing the refined structures yield ³⁵Cl EFG tensors that agree well with experiment. PXRD and ³⁵Cl SSNMR, in tandem with reliable calculations of EFG tensors, are essential for the development of NMR crystallographic and crystal structure prediction protocols and crucial for future studies involving HCl salts and their concomitant solid forms.

1. INTRODUCTION

The physicochemical properties of active pharmaceutical ingredients (APIs), including stability, solubility, bioavailability, and reactivity, depend on the structures of their solid forms,^{1–12} which include salts, polymorphs, hydrates/solvates, cocrystals, amorphous solid dispersions, and combinations thereof.¹³⁻¹⁶ The synthesis of polymorphs (*i.e.*, different crystalline forms containing the same molecular components), salts (i.e., solids exhibiting complete charge transfer between two or more components), hydrates/solvates (i.e., solids in which water/solvent molecules are incorporated into the crystal lattice), and/or cocrystals (i.e., solids that are composed of two or more molecular and/or ionic components in the same crystal lattice, which are neither solvates nor simple salts) affords additional opportunities for enhancing desired physicochemical properties of pharmaceutical products. Given the importance of the production of solid forms of APIs to the pharmaceutical industry, the propensity of neutral organic molecules and their salts to exhibit polymorphism, as well as cocrystal and hydrate/solvate formation, has been evaluated through mining of databases and quantum chemical

calculations.^{17–20} Furthermore, the potential for tailoring the physicochemical properties of APIs through the production of a variety of different solid forms is crucial for the design and manufacture of dosage formulations of high efficacy, and may even provide insight into the rational design of new solid forms.

Intermolecular noncovalent interactions in solid APIs that influence their structure and concomitant physicochemical properties are of great interest. In particular, understanding H…Cl⁻ hydrogen bonding is very important since nearly half of all commercial pharmaceuticals are constituted by hydrochloride (HCl) salts of APIs.²¹ In HCl salts, H…Cl⁻ hydrogenbonding interactions are a major source of structural stability

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since the chloride ions can accommodate multiple hydrogen bonds, leading to a wide variety of structural arrangements. Furthermore, the presence of solvent molecules (most commonly water) and/or coformer molecules (in the case of cocrystals) can impact the properties of HCl APIs through the formation of new types of H…Cl⁻ hydrogen bonds. Therefore, an understanding of H…Cl⁻ hydrogen-bonding interactions allows for informed choice of solvents and coformers, which is paramount for the rational design of new solid forms with specifically tailored physicochemical properties.

Key to the successful production of polymorphs, hydrates, and cocrystals of APIs is a reliable and rapid means of molecular-level structural characterization. Characterization methods such as powder and single-crystal X-ray diffraction (PXRD and SCXRD, respectively) thermal analysis, vibrational spectroscopy (IR and Raman), and solid-state NMR (SSNMR) spectroscopy all play important roles in this respect.^{22,23} Of these, SSNMR spectroscopy is the premier technique for providing atomic-level structural detail. Onedimensional (1D) and two-dimensional (2D) ¹³C, ¹⁵N, ¹⁹F, and ¹H SSNMR experiments are commonly used to characterize the molecular structures of APIs and are well suited for exploring noncovalent interactions that are key in determining the arrangements of the molecules in different solid forms.^{23–29}

Our group and others have demonstrated that ³⁵Cl SSNMR spectroscopy is invaluable for rapid fingerprinting and differentiation of different solid forms of the APIs in both bulk and dosage forms, as well as probing the local geometries of ions, including the types, numbers, and arrangements of chemical species participating in H…Cl- hydrogen bonding.³⁰⁻⁵⁷ The ³⁵Cl SSNMR spectra of chloride ions in organic HCl salts almost always feature broad central-transition (CT, $+1/2 \leftrightarrow -1/2$) powder patterns that are influenced by the second-order quadrupolar interaction (SOQI), and in many cases, chlorine chemical shift anisotropy (CSA). The ³⁵Cl electric field gradient (EFG) tensor, which gives rise to the QI, is extremely sensitive to changes and/or differences in chloride ion environments that result from distinct hydrogen-bonding arrangements. Different solid forms of APIs have distinct arrangements of H…Cl⁻ hydrogen bonds, with hydrates and cocrystals even featuring additional types of hydrogen bonds involving water molecules or coformers, respectively. Accordingly, ³⁵Cl SSNMR spectroscopy is a robust tool for the structural characterization of solid forms of APIs, including their polymorphs, hydrates, and cocrystals.

Xylazine HCl, (2-(2,6-xylidino)-5,6-dihydro-4H-1,3-thiazine hydrochloride), denoted here as X, is used as a sedative, analgesic, and muscle relaxant in veterinary medicine (Scheme 1).⁵⁸ There are five reported solid forms of X that can be produced under ambient conditions, including one hydrate (Xh) and four anhydrous polymorphs (Xx, Xa, Xz, and Xm), the crystal structures of which have been reported for all forms except Xm.⁵⁹⁻⁶¹ There are additional hydrates and solvates

Scheme 1. Molecular Structure of Xylazine HCl



that can be made under conditions of high pressure;⁶¹ however, these will not be discussed in the current study. There are reports that the anhydrous forms of X slowly convert into **Xh** under ambient conditions, and can rapidly convert into **Xh** under controlled environmental conditions (*i.e.*, hydration chamber or recrystallization from water).⁶¹ Furthermore, **Xh** can be used as a reagent in the synthesis of the anhydrous polymorphs **Xx**, **Xa**, and **Xz** under various synthetic conditions (*i.e.*, heating under vacuum, recrystallization, *etc.*).^{59,62,63} This complex array of interconversions between the reported hydrated and anhydrous forms (Scheme 2) makes these systems ideal for a case study of polymorph discovery and/or interconversion, hydration state identification, as well as for exploring accelerated mechanochemical interconversions and cocrystal design.

Herein, we discuss the synthesis, characterization, and structural refinement of a monohydrate, three known polymorphs, one novel polymorph, and three novel cocrystals of X (Table 1). In all cases, solid forms of X are characterized with a combination of PXRD, SCXRD, DSC, TGA, and multinuclear SSNMR spectroscopy (¹³C and ³⁵Cl). First, we discuss the reported and novel mechanochemical syntheses of Xh, Xx, Xa, and Xz, and demonstrate specific pathways for their interconversion. Second, utilizing information garnered from similar crystal systems and accompanying PXRD and ³⁵Cl SSNMR data, ball milling protocols are used to synthesize the new polymorph Xy and three cocrystals of X with benzoic acid (Bzc) and benzamide (Bza) in 1:1 ratios (**XBzc** and **XBza**) and malonic acid (Mal) in a 2:1 ratio (X₂Mal). We also report crystal structures from SCXRD for the novel polymorph and cocrystals. ³⁵Cl SSNMR spectroscopy provides a unique spectral fingerprint for each solid form of X (and cocrystals), which is associated with unique sets of chlorine EFG and chemical shift (CS) tensors. Third, dispersion-corrected planewave DFT (DFT-D2*) methods are used to refine all crystal structures, with the validity of the structures assessed via comparison of experimental and calculated ³⁵Cl EFG tensors. Relationships between the local chloride hydrogen-bonding environments and the ³⁵Cl EFG tensor parameters and orientations are also investigated. Finally, the future application of quadrupolar NMR-based crystallography and crystal structure prediction of APIs, polymorphs, hydrates, and cocrystals is discussed briefly.

2. EXPERIMENTAL SECTION

2.1. Materials. Samples of X, Bzc, and Mal were purchased from Millipore Sigma. Bza was purchased from Oakwood Chemical. PXRD indicates that different batches of the as-received samples of X consist either of pure phases of Xz or Xa, or as a mixture of Xz and Xh (*vide infra*). All other reagents consist of pure phases. Solvents, including methanol (MeOH), ethanol (EtOH), acetonitrile (MeCN), dimethylformamide (DMF), and dichloromethane (DCM), were purchased from VWR. All powder samples were stored in 20 mL glass vials, sealed with plastic screw caps, and stored under conditions of controlled relative humidity (except for Xx, which was transferred and packed in a glovebox to minimize contact with air/water).

2.2. Mechanochemical Synthesis. All ball mill syntheses involved liquid-assisted grinding (LAG) or neat grinding (NG) using a Retsch Mixer Mill 400 at a milling frequency of 20 Hz for 90 min. All educts were placed in a vacuum oven overnight prior to their usage to minimize surface water absorption. The materials were placed in a 10 mL Teflon milling jar with two 5 mm stainless steel ball bearings. Materials used for LAG synthesis were added to the milling jars prior to the addition of solvent. Parameters used in the mechanochemical syntheses can be found in Tables S1 and S2

Scheme 2. Summary of Previously Reported Reaction Pathways for the Synthesis of Xylazine HCl (X) Anhydrous Polymorphs and the Monohydrate: (i) Formation of Xh through Slow Recrystallization of Xa, Xm, Xx, or Xz from Water or Placement in Hydration Chamber; (ii) Heating at (a) 100 °C for 2 Days and (b) 60 °C for 1 Day in a Vacuum Oven, to Synthesize Xa and Xx, Respectively; (iii) Recrystallization of Xa from Methanol to Obtain Xz, or Recrystallization from DCM to Form Xm (Following the Desolvation of an Intermediate Phase)



Table 1. Summary of Solid Forms of Xylazine HCl, Abbreviated Names Used in This Work, and an Overview of Crystallographic Data

form of xylazine HCl	abbreviation	space group	Z, Z'	refs
monohydrate	Xh	$P2_1/c$	4, 1	this work
form A	Xa	$P2_1/c$	4, 1	59
form X	Xx	$P2_{1}/c$	4, 1	59
form Z	Xz	$P2_1/c$	4, 1	59
form M	Xm	unknown	unknown	n/a
form Y	Ху	$P6_1$	6, 1	this work
benzoic acid 1:1 cocrystal	XBzc	C2/c	8, 1	this work
benzamide 1:1 cocrystal	XBza	$P\overline{1}$	2, 1	this work
malonic acid 2:1 cocrystal	X ₂ Mal	P2 ₁ 2 ₁ 2 ₁	4, 1	this work

(Supporting Information). Mechanochemical interconversions between the different polymorphs and hydrate of X were all completed by LAG methods. The starting material and solvent were placed in Teflon jars and milled in the presence of two 5 mm stainless steel or 3 Å 4–8 mesh molecular sieves as ball bearings. Syntheses of the **XBzc**, **XBza**, and **X**₂**Mal** PCCs were completed by NG, in a 1:1 ratio using any form of X and coformers Bza or Bzc, or a 2:1 ratio of X and Mal. No further grinding of samples was done prior to packing into plate sample holders for analysis by PXRD or glass tubes/zirconia rotors for analysis by SSNMR.

2.3. Powder X-ray Diffraction. PXRD patterns for all samples were acquired using a Proto AXRD benchtop diffractometer with a Cu K α radiation source and a DECTRIS hybrid pixel detector using the XRDWin software. Samples were mounted on plate sample holders manufactured by Proto. The X-ray tube voltage and current were 30 kV and 20 mA, respectively. The divergence and scattering slits were set to 0.5 and 0.6 mm, respectively. Diffraction experiments were run with the detector scanning a 2θ range from 5 to 45° implementing a dwell time of 10 seconds, resulting in ca. 55 min of acquisition time per sample. Additional confirmations of sample identities and purities were obtained via PXRD experiments conducted using a Rigaku Miniflex benchtop diffractometer with a Cu K α radiation source and D/tex Ultra silicon strip detector. The Xray tube voltage and current were set to 40 kV and 15 mA, respectively. All samples were packed in zero-background silicon wafers with a well size of 5 mm × 0.2 mm mounted on an eightposition autosampler. Diffraction experiments were run with the detector scanning 2θ angles from 5 to 50° with a step size of 0.03° and speed of 5° min⁻¹ (acquisition time of *ca*. 10 min). The resulting PXRD patterns for all samples were compared to single-crystal simulations for purity assessment and identification of starting reagents and final products, using the CrystalDiffract software package.

2.4. Single-Crystal X-ray Diffraction. Crystals were coated in paratone-N oil and mounted onto a goniometer head using a polyimide cryoloop. The crystals were mounted and cooled to 170 \pm 2 K using an Oxford Cryosystems Cryostat (700 Series Cryostream Plus) attached to the diffractometer. X-ray reflections were collected and integrated from frame data using the APEX-III⁶⁴ software suite on a Bruker D8 Venture four-circle diffractometer equipped with a Photon CCD detector utilizing a graphite monochromator with either Cu K α (λ = 1.54178 Å) or Mo K α radiation (λ = 0.71073 Å). The SAINT⁶⁴ software package was used for raw area detector data frame reductions and SADABS⁶⁴ was used for absorption corrections (multiscan; semiempirical from equivalents). Structures were solved using intrinsic phasing⁶⁵ and refined against F^2 with SHELXL⁶⁶ software implemented within the OLEX2⁶⁷ program. The refinement of all nonhydrogen atoms was completed anisotropically. In all structures, hydrogens attached to N2 and N4 were identified in difference electron density maps, while all other hydrogen atoms were located in idealized positions and refined using a riding model. The structures have been deposited on the Cambridge Structural Database with CSD deposition numbers 2214351-2214355.

2.5. Thermal Analysis. Thermogravimetric analysis (TGA) studies were conducted using a TA Instruments TGA5500. Differential scanning calorimetry (DSC) studies were conducted using a TA Instruments DSC2500. N₂ gas with 99.999% purity was used to purge the systems at a flow rate of 60 mL min⁻¹. Samples (*ca.* 5 mg) were placed in aluminum pans and referenced against an empty pan for the DSC measurements. TGA samples were held at 25 °C for 5 min before heating to 500 °C at a rate of 5 °C min⁻¹.

2.6. Solid-State Nuclear Magnetic Resonance Spectroscopy. 2.6.1. Overview. All moderate-field SSNMR experiments were conducted at the University of Windsor (Windsor, Ontario) and acquired using a Bruker Avance III HD NMR spectrometer with an Oxford wide-bore magnet with resonance frequencies of $\nu_0(^{13}\text{C}) =$ 100.64 MHz, $\nu_0({}^{1}\text{H}) = 400.224$ MHz, and $\nu_0({}^{35}\text{Cl}) = 39.21$ MHz. Static experiments were conducted using a revolution 5 mm HX ultralow temperature stationary probe with samples packed into 5 mm outer diameter (o.d.) glass tubes sealed with Teflon tape. Magic-angle spinning (MAS) NMR experiments were conducted using a Varian/ Chemagnetics 4 mm HX MAS probe with samples packed into 4 mm o.d. zirconia rotors with a Teflon cap. All high-field SSNMR experiments were conducted at the National High Magnetic Field Laboratory (Tallahassee, Florida). Samples of Xz, Xx, Xa, and Xh were packed into 3.2 mm o.d. zirconia rotors and acquired under static and MAS conditions using a Bruker Avance III HD spectrometer, a 31 mm bore Oxford magnet, and a home-built 3.2 mm HX MAS probe $(B_0 = 19.5 \text{ T}, \nu_0)^{(1)} = 831.48 \text{ MHz}$, and $\nu_0({}^{35}\text{Cl}) = 81.46 \text{ MHz}$). Samples of **XBza**, **XBzc**, **X**₂**Mal**, and **Xy** were

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material		$C_{\rm Q}$ (MHz)	$\eta_{\rm Q}$	$\delta_{ m iso}~(m ppm)$	Ω (ppm)	κ	α (deg.)	β (deg.)	γ (deg.)
Xh	exp.	3.59(6)	0.80(4)	76(4)	65(10)	0.0(4)	80(10)	75(5)	5(10)
	RPBE-D2*	3.80	0.53	80	62	-0.27	99	82	8
	SCXRD	2.53	0.53	49	41	-0.31	101	81	18
Xa	exp.	3.29(6)	0.87(4)	131(5)	110(30)	0.5(4)	0(10)	10(10)	95(5)
	RPBE-D2*	-3.56	0.63	141	132	0.87	168	5	124
	PXRD	2.70	0.92	128	134	0.49	109	89	12
Xx	exp.	5.28(6)	0.28(4)	79(5)	150(20)	0.1(2)	0(10)	5(10)	100(5)
	RPBE-D2*	-4.72	0.50	76	148	-0.32	101	4	13
	PXRD	2.90	0.88	71	121	-0.63	100	90	4
Ху	exp.	6.15(8)	0.14(5)	79(5)	120(50)	f	f	0(30)	f
	RPBE-D2*	-6.08	0.17	95	148	0.71	65	4	16
	SCXRD	-2.75	0.64	68	106	0.73	5	5	37
Xz	exp.	3.94(5)	0.64(4)	80(4)	80(30)	-0.7(2)	165(10)	90(10)	0(30)
	RPBE-D2*	-3.40	0.99	87	120	-0.64	161	9	76
	PXRD	-2.92	0.93	86	105	-0.69	55	17	48
XBzc	exp.	5.23(6)	0.50(4)	107(5)	140(30)	0.0(4)	20(10)	25(10)	95(10)
	RPBE-D2*	-5.22	0.68	112	144	-0.03	15	30	98
	SCXRD	-2.57	0.07	78	111	-0.06	41	29	106
XBza	exp.	5.35(5)	0.51(3)	91(4)	100(30)	f	f	0(10)	f
	RPBE-D2*	-5.06	0.60	89	96	0.15	33	9	121
	SCXRD	-2.32	0.63	57	65	-0.15	52	18	34
X ₂ Mal (Cl1)	exp.	4.28(8)	0.30(5)	92(5)	50(30)	f	f	0(10)	f
	RPBE-D2*	3.98	0.35	109	47	-0.10	147	69	30
	SCXRD	3.07	0.66	79	34	-0.86	141	69	117
X ₂ Mal (Cl2)	exp.	4.73(8)	0.97(5)	127(5)	100(50)	f	85(20)	85(20)	f
	RPBE-D2*	4.93	0.91	144	95	-0.09	115	88	24
	SCXRD	2.13	0.44	99	43	-0.22	116	87	41

Table 2. Experimental and Calculated ³⁵Cl EFG and Chemical Shift Tensor Parameters of Xylazine HCl Polymorphs, Hydrates, and Cocrystals^{*a,b,c,d,e*}

^{*a*}Theoretical EFG and CS tensor parameters were obtained from calculations on XRD-derived structures and structures refined at the RPBE-D2* level. ^{*b*}The experimental uncertainties in the last digit for each value are indicated in parentheses. ^{*c*}The principal components of the EFG tensors are defined such that $|V_{33}| \ge |V_{22}| \ge |V_{11}|$. The quadrupolar coupling constant and asymmetry parameter are given by $C_Q = eQV_{33}/h$ and $\eta_Q = (V_{11} - V_{22})/V_{33}$, respectively. The sign of C_Q cannot be determined from the experimental ³⁵Cl spectra. ^{*d*}The chemical shift tensors are defined using the Herzfeld–Berger convention, in which the principal components are ordered $\delta_{11} \ge \delta_{22} \ge \delta_{33}$. The isotropic chemical shift, span, and skew are given by $\delta_{iso} = (\delta_{11} + \delta_{22} + \delta_{33})/3$, $\Omega = \delta_{11} - \delta_{33}$, and $\kappa = 3(\delta_{22} - \delta_{iso})/\Omega$, respectively. ^{*e*}The Euler angles α , β , and γ define the relative orientation of the EFG and magnetic shielding tensors using the ZY'Z'' convention for rotation, in which the principal values of the magnetic shielding tensor are ordered $\sigma_{11} \le \sigma_{22} \le \sigma_{33}$. The experimental angles derived from ssNake (which uses the ZX'Z'' convention) are adjusted to match the calculated values extracted by EFGShield according to the following operations: $\alpha = \gamma(ssNake)$; $\beta = \beta(ssNake)$; $\gamma = \alpha(ssNake) + 90^{\circ}$. ^{*f*}This parameter has little to no effect on the simulated ³⁵Cl SSNMR pattern.

packed into 3.2 or 1.3 mm o.d. zirconia rotors and acquired under static and MAS conditions using a Bruker NEO spectrometer, an 89 mm bore Oxford magnet, and either a home-built 3.2 mm HXYD MAS probe or a home-built 1.3 mm HXY MAS probe ($B_0 = 18.8$ T, $\nu_0(^{1}\text{H}) = 800.13$ MHz, and $\nu_0(^{35}\text{Cl}) = 78.36$ MHz). Full details of the experimental parameters are provided in the Supporting Information (Tables S3–S9).

2.6.2. ¹³*C* SSNMR Spectra. ¹³*C* SSNMR spectra of all forms of X were acquired at 9.4 T using a ${}^{1}\text{H} \rightarrow {}^{13}\text{C}$ variable-amplitude crosspolarization (VACP) MAS technique, ${}^{68-71}$ using SPINAL-64 ${}^{1}\text{H}$ decoupling with $\nu_2({}^{1}\text{H}) = 50$ kHz, a spinning rate of $\nu_{rot} = 11$ or 12 kHz, a $\pi/2$ ${}^{1}\text{H}$ pulse width of 5.0 μ s, optimized contact times between 0.7 and 0.9 ms, and an optimized recycle delay of 10 s. Spectra for the coformers were collected using a $\pi/2$ ${}^{1}\text{H}$ pulse width of 2.0 μ s, optimized contact times between 1.0 and 1.7 ms, and a recycle delay of 250 s. Chemical shifts were referenced to TMS using the carbonyl carbon in ${}^{13}\text{C}$ -labeled α -glycine at $\delta_{iso}({}^{13}\text{C}) = 176.5$ ppm as a secondary reference.

2.6.3. ³⁵Cl SSNMR Spectra. Static ³⁵Cl{¹H} SSNMR spectra were acquired using Hahn-echo, ⁷⁴ CPMG, ⁷⁵ or WURST-CPMG⁷⁶⁻⁷⁹ pulse sequences with continuous wave (CW) ¹H decoupling at $\nu_2(^{1}H) = 25-35$ kHz. ³⁵Cl{¹H} MAS SSNMR spectra ($\nu_{rot} = 16$ or 30 kHz) were acquired using the Hahn-echo sequence, sometimes utilizing rotor-assisted population transfer (RAPT) via 64 μ s WURST-80

pulses,^{80,81} and a CW ¹H decoupling field of $\nu_2(^{1}\text{H}) = 25$ or 55 kHz. Chemical shifts were referenced to NaCl(s) at $\delta_{iso} = 0.0$ ppm. All spectra were processed and fit using the ssNake software package.⁸² Uncertainties in quadrupolar and chemical shift parameters were estimated *via* bidirectional variation of the parameters, and visual comparison of various simulations overlaid with spectra. The reported Euler angles from ssNake, which describe the relative orientation of the magnetic shielding and EFG tensors, are converted from the *ZX'Z*" convention for rotation to the *ZY'Z*" convention, to facilitate comparison with calculated values (see Table 2 for details).^{82,83}

2.7. Plane-Wave Density Functional Theory (DFT) Calculations. All plane-wave DFT calculations were performed using the CASTEP module within Materials Studio 2020.⁸⁴ The calculations employed the RPBE functional,⁸⁵ a plane-wave cutoff energy of 800 eV, and ZORA ultrasoft pseudopotentials generated on the fly.⁸⁶ Integrals over the Brillouin zone were sampled using a Monkhorst–Pack grid with a *k*-point spacing of 0.05 Å^{-1.87} Dispersion was implemented in the geometry optimizations using a modification of Grimme's two-body dispersion force field,⁸⁸ as discussed in previous work.^{89–91} The SCF convergence threshold was 5×10^{-7} eV atom⁻¹. Geometry optimization and energy minimization used the lowmemory BFGS algorithm.⁹² Structural convergence was assessed using a maximum change in energy of 5×10^{-6} eV atom⁻¹, a maximum displacement of 5×10^{-4} Å atom⁻¹, and a maximum



Figure 1. Experimental ³⁵Cl^{{1}H} SSNMR spectra of **Xh** (blue), **Xa** (red), **Xx** (orange), and **Xz** (purple), along with corresponding analytical simulations (black). Spectra at $B_0 = 9.4$ T were acquired under static conditions with the WURST-CPMG pulse sequence. Spectra at $B_0 = 19.5$ T were acquired under static and MAS ($\nu_{rot} = 16$ kHz) conditions using the Hahn-echo pulse sequence. Spinning sidebands are indicated with a sterisks (*). Impurity arising from **Xh** is denoted with a dagger (†).

Cartesian force of 10^{-2} eV Å⁻¹. Chlorine magnetic shielding tensors were computed using the GIPAW approach⁹³ and converted to the chemical shift scale relative to solid NaCl by setting the calculated magnetic shielding of NH₄Cl to $\delta_{iso}(^{35}Cl) = 120$ ppm.⁹⁴ Euler angles, which use the ZY'Z" convention for rotation, were extracted from the CASTEP output files using EFGShield 4.1.⁸³ The procedure for calculating ¹³C chemical shifts uses both CASTEP and the Amsterdam Modelling Suite (AMS), as described in Supporting S1.

3. RESULTS AND DISCUSSION

3.1. Overview and General Observations. 3.1.1. Overview. We discuss optimized conditions required for the successful synthesis and/or interconversion of four anhydrous polymorphs and one monohydrate of X; this includes a discussion of a novel polymorph, Xy, whose synthesis and structural characterization are presented here for the first time. In addition, we discuss the production and characterization of three novel cocrystals of X via ball milling. Ball milling conditions (frequency, number of ball bearings, and milling and ball bearing materials) and solvents were chosen to optimize for maximum yields for all preparations. PXRD, thermal analysis, and ¹³C and ³⁵Cl SSNMR data, along with structural interpretations, are presented to confirm the identities of the products and potential impurities. Crystal structures from SCXRD are reported for the four novel solid forms, and the structure of Xh is redetermined.

Before proceeding to a discussion of each system, general observations on features from PXRD, and ^{13}C and ^{35}Cl SSNMR data are discussed. Then, detailed descriptions of the

syntheses, thermal analysis, SCXRD, ¹³C SSNMR, and ³⁵Cl SSNMR characterizations of four anhydrous polymorphs and one monohydrate of X are presented, followed by the three novel cocrystals of X. We discuss the refinement of the crystal structures of these eight forms of X using dispersion-corrected plane-wave DFT calculations (*i.e.*, DFT-D2*).^{89–91} Finally, we present DFT calculations of ³⁵Cl EFG tensors and discuss their relationships with the local structural environments of the chloride ions, which may aid future attempts to predict structures of novel solid forms of X, or other APIs and associated cocrystals, in the absence of diffraction data.

3.1.2. Powder X-ray Diffraction. Prior to the synthesis of different solid forms of X and its cocrystals, PXRD patterns were acquired for all solid reagents. The PXRD patterns of different "as-received" samples of X reveal either Xa, Xz, or a mixture of Xh and Xz (Figure S1), based on comparisons to simulated PXRD patterns from the known crystal structures (this raised questions about the best starting point for developing an understanding of interconversions among the different solid forms, vide infra).⁶¹ PXRD patterns of Bzc, Bza, and Mal (Figure S2) match well with those simulated from known SCXRD data.^{59–61} The PXRD patterns of the cocrystals indicate pure products and do not reveal any evidence of starting materials (Figures S3–S5).

In part, PXRD data support the successful synthesis of the different solid forms of X and its cocrystals (*vide infra*). PXRD patterns of all previously reported solid forms of X match well with simulations and indicate samples of high crystallinity

crystal structures (Figure S7). Finally, because the samples were prepared over a period of several years, and in multiple locations with different atmospheric conditions, PXRD patterns were acquired prior to analysis by SSNMR, to provide an additional avenue for characterizing the materials and assessing their purities.

3.1.3. ${}^{1}H \rightarrow {}^{13}C$ SSNMR Spectra. ${}^{13}C$ CP/MAS spectra acquired at 9.4 or 14.1 T are useful for phase characterization, detection of impurities, and basic structural interpretation. The detection of impurity phases arising from incomplete transformations from the starting educt of X and/or coformers, or the interconversion between phases due to exposure to atmospheric conditions, provides a complementary method to PXRD for characterizing samples. Additionally, peak assignments can be made with the aid of DFT calculations (Tables S10 and S11), which can provide useful site-specific chemical interpretations (*vide infra*).

3.1.4. ³⁵Cl SSNMR Spectra. ³⁵Cl^{{1}H} MAS NMR spectra were acquired at 18.8 or 19.5 T to separate the central isotropic powder pattern from the spinning sidebands, allowing for the determination of the quadrupolar coupling constants, C_{Q_2} asymmetry parameters, η_{Q_2} and isotropic chemical shifts, δ_{iso} . Static ³⁵Cl^{{1}H} SSNMR spectra were collected at 9.4 T, and either 18.8 or 19.5 T, in order to deconvolute spectral contributions from the EFG and CS tensors, to refine the parameters obtained from MAS NMR spectra, and to extract the anisotropic CS parameters and Euler angles describing the relative orientation of the EFG and CS tensors (see Table 2 for a complete listing of all EFG and CS tensor parameters and conventions).

The ³⁵Cl¹H} SSNMR spectra of all samples are typical of those of chloride ions in HCl salts of organic APIs (Figures 1-3). Most of the CT patterns are dominated by contributions from the SOQI, though the effects of chlorine CSA can be observed in the high-field spectra (N.B.: because of the MAS NMR spectra, the uncertainties in δ_{iso} , C_Q , and η_Q are small in comparison to those of the CS tensor parameters and Euler angles). The difference between each of the patterns is visually striking, with each having key discontinuities that arise from the unique EFG and CS tensors and their distinct relative orientations. In all cases, it is clear that these spectra correspond to very different chloride ion environments, even without recourse to simulations, meaning that the ³⁵Cl CT powder patterns function as useful spectral fingerprints for rapidly and reliably distinguishing solid forms. The ${}^{35}Cl{}^{1}H$ SSNMR spectra for all materials feature a number of CT patterns that are consistent with the number of crystallographically distinct chloride ions indicated by the crystal structures. ³⁵Cl SSNMR spectra are very useful for the detection of impurities, which are often not apparent in corresponding PXRD patterns. More details on the ³⁵Cl SSNMR spectra of each sample, the anisotropic NMR interaction parameters derived from these spectra, and their relationships to synthesis and structure, are discussed below.

3.2. Hydrate and Anhydrous Polymorphs. *3.2.1. Synthesis and Characterization.* There are five previously reported solid forms of X under ambient conditions: one

monohydrate (Xh) and four anhydrous polymorphs (Xa, Xx, Xz, and Xm), ^{59,61} with reported preparations shown in Scheme 2. Form Xh can be produced from Xa, Xx, or Xz by slow recrystallization from water or placement in a hydration chamber. Conversely, form Xh can be heated at 100 °C for 2 days to produce Xa, or to 48-60 °C for 1 day to produce Xx. Interestingly, Xz and Xm are not reported to be formed in this manner; instead, Xa is converted to Xz through recrystallization from dry MeOH,^{62,63} whereas Xm results from the desolvation of the DCM solvate of X (which is prepared by recrystallization of Xa from DCM).⁶³ Several studies have examined the relative stabilities of these solid forms, as well as the kinetics of their phase transitions. Xh is the most stable phase,⁶³ whereas the relative stabilities of the anhydrous polymorphs vary with temperature.⁶² Of the anhydrous polymorphs, Xz is the most stable near RT, whereas Xa is the most stable above 48 $^\circ$ C, and Xx is the least stable across this temperature range. The transformation of Xx to Xa has been explored in some detail,⁹⁵ as has the dehydration of **Xh** to produce Xx or Xa.⁹⁶ Factors that are known to influence the kinetics of phase transformations include temperature, pressure, humidity, and particle size distribution of the educt.

We found commercial samples of as-received X to contain different solid forms (*i.e.*, **Xa**, **Xz**, or a mixture of **Xz** and **Xh**, Figure S1). This led us to carefully investigate the rigid sets of synthetic conditions to reliably produce a single solid form. We found that no matter what the initial starting form is, we can reliably produce **Xh** *via* (i) recrystallization from distilled water by slow evaporation;⁶² (ii) storing the sample in a hydration chamber for *ca.* 2 days;⁶³ or (iii) ball milling the sample in the presence of 0.5 mol equiv of water for 30 min (this latter method has not been reported to date). Additionally, we were able to grow single crystals of **Xh** by slow evaporation from distilled water; from these, we redetermined the structure *via* SCXRD (*vide infra*).

Once we established the robust production of pure Xh, we initiated a series of experiments aimed at producing the other solid forms. First, attempts to produce the anhydrous polymorphs Xa and Xx by dehydration of Xh under vacuum using previously reported procedures were successful, with complete dehydration observed at 100 and 50 °C, respectively, after 3 days.⁹⁶ Attempts to prepare Xa and Xx by ball milling under ambient conditions were unsuccessful, as they only seem to form under conditions of low pressure (N.B.: now that we could reliably produce the stable forms Xa and Xx, we also used these as educts for the synthesis of new solid forms). Second, we found that Xz, which can be prepared via recrystallization of Xa from MeOH,62 could also be produced by LAG of Xa, Xx, Xz, or Xh with two 5 mm stainless steel or 3 Å 4-8 mesh molecular sieves in solvents such as MeOH, EtOH, or DMF. The molecular sieves were found to be crucial for trapping water and preventing the persistence and/or formation of **Xh** (or other potential hydrated forms) within the Teflon-based milling containers. Third, we attempted to make Xm, which was reported as being prepared by a two-step process involving (i) recrystallization of Xa from DCM to yield a DCM solvate of X and (ii) subsequent desolvation over one day at low RH to yield \mathbf{Xm} ;⁶³ our attempts at duplicating this result were unsuccessful. However, after numerous trials involving LAG with any form of X, two molecular sieves, and MeCN, we produced a novel anhydrous polymorph, Xy. We note that milling Xa with MeCN and two stainless steel ball bearings, rather than molecular sieves, led to incomplete





LAG using two molecular sieves and acetonitrile





Figure 2. Experimental (green) and simulated (black) 35 Cl{ 1 H} static and MAS SSNMR spectra of novel anhydrous form **Xy** acquired at 9.4 and 18.8 T. Static and MAS spectra reveal the presence of an impurity, **Xh**, from hydration, indicated in the simulation in light blue. Spinning sidebands are indicated with asterisks (*).

conversion to Xy, resulting in a mixture of polymorphs easily observable in ³⁵Cl SSNMR (Figure S15).

Previous reports of interconversions between solid forms of X, as well as the novel LAG experiments described above, convinced us to further explore the landscape of possible synthetic conditions. Therefore, we took each of the anhydrous solid forms and subjected them to the synthetic conditions described above (see Scheme 3). We found that in every case, it was possible to produce each solid form in high yields (*i.e.*, with little or no impurity detected by PXRD), provided that the synthetic conditions were rigorously adhered to. Hence, it appears that the formation of each solid form of X is independent of the educt and dependent solely on the synthetic conditions.

3.2.2. Thermal Analysis. DSC thermograms and TGA curves for the monohydrate and anhydrous polymorphs of X are found in Figures S8 and S9. The DSC thermograms for Xa and Xy feature a single endothermic peak with no loss of mass observed in the corresponding TGA curves, which likely corresponds to the melting of the solid. In contrast, the DSC thermograms for Xx and Xz feature multiple endothermic events. Of these, the events in the range of *ca.* 100–110 °C likely correspond to the disproportionation of the HCl salts (which is suggested by the loss of mass in the TGA curves). A second event corresponding to the melting of freebase X is observed at *ca.* 140 °C.⁹⁷ Possible solvent loss is also observed. Finally, the DSC thermogram for Xh exhibits two endothermic

events, neither of which coincide with mass loss in the TGA curve, suggesting the formation of a new phase at *ca.* 80–90 °C, before melting. Previous work has illustrated that the dehydration of Xh yields either Xa or Xx, depending on conditions such as temperature, pressure, and humidity;⁶³ our results indicate a single melting endotherm, illustrating that dehydration of Xh yields Xa uniquely, under these experimental conditions.

3.2.3. Crystal Structures. The SCXRD structures of Xa, Xx, Xz, and Xh have been reported previously.⁵⁹ We report a new crystal structure for the monohydrate Xh (Figure S16), which differs from the previously reported structure acquired at different temperatures and pressures,^{59,61} as well as that of a novel anhydrous polymorph, Xy (Figure S17). The previously reported anhydrous polymorphs and monohydrate of X all belong to the monoclinic $P2_1/c$ space group (no. 14, Z = 4, Z' = 1), whereas the novel form Xy crystallizes in the hexagonal space group $P6_1$ (no. 169, Z = 6, Z' = 1). It is also noteworthy that the molecular-level structure of the xylazine molecule within Xy differs significantly from that of all previously reported solid forms of X; this is most clearly reflected in the N3–C4–N5–C6 torsional angle, which is *ca.* 5° in Xy, but closer to *ca.* 175° for the other forms.

3.2.4. ${}^{1}H \rightarrow {}^{13}C$ SSNMR Spectra. The ${}^{1}H \rightarrow {}^{13}C{}^{1}H$ VACP/ MAS spectra of the anhydrous polymorphs and monohydrate of X have common groupings of chemical shifts in regions between *ca*. 10–50 and *ca*. 120–170 ppm (Figure S10). Slight



Figure 3. Experimental ³⁵Cl{¹H} SSNMR spectra of **XBzc** (red), **XBza** (green), and **X₂Mal** (purple) along with the corresponding analytical simulations (black). Spectra at $B_0 = 9.4$ T under static conditions were acquired with the WURST-CPMG pulse sequence. Spectra at $B_0 = 18.8$ T under static and MAS ($\nu_{rot} = 30$ kHz) conditions were acquired using the QCPMG and RAPT-echo pulse sequences, respectively. Spinning sidebands are indicated with asterisks (*).

differences in chemical shifts among spectra are expected for the distinct polymorphs and hydrated phases. In each case, several of the peaks overlap (especially in the methyl and aromatic regions), preventing the determination of the chemical shifts for all twelve carbon atoms within the xylazine molecule and concomitant crystal lattice. Furthermore, the spectra of Xa, Xx, and Xy all feature additional peaks that we attribute to the presence of impurity phases of X. In the cases of Xa, the impurity corresponds only to Xz, whereas for Xa and Xy, the impurities are attributable to both Xz and Xh. We note that these impurity phases were not always detected beforehand by PXRD (i.e., no impurity was detected in the PXRD pattern for Xa, whereas the pattern for Xx indicated only the Xh impurity); this suggests that the samples sometimes contain impurity phases that are below the limit of detection of PXRD due to incomplete transformation from the educt form of X, as part of an ongoing transformation process due to exposure to atmospheric water, or even that the transformations between solid forms of X could be facilitated in situ by frictional heating during magic-angle spinning. DFT calculations of the ¹³C chemical shifts are illustrated in Figure S11; however, the authors stress that peak assignments made on the basis of isotropic chemical shifts alone often result in errors, but can be facilitated through various 2D methods.⁹

3.2.5. ³⁵Cl SSNMR Spectra. ³⁵Cl{¹H} CT patterns acquired at 9.4 T range in breadth from *ca*. 70–150 kHz, while those acquired at 18.8 or 19.5 T are narrower for both static and MAS experiments (*ca*. 33–78 and *ca*. 22–26 kHz, respectively) due to the inverse field dependence of the SOQI on pattern breadth (Figures 1 and 2). These pattern breadths are typical for CT powder patterns of chloride ions in HCl salts.^{30,31,36} Simulations of the ³⁵Cl{¹H} spectra acquired at multiple fields for all forms of X allow for the determination of the EFG and CS tensor parameters, with the magnitudes of $C_{\rm Q}$ ranging from 3.29 to 6.15 MHz (Table 2) and Xy having the largest.

³⁵Cl SSNMR spectra are very useful for the detection of impurities, which were only detected in the spectra of Xz and Xy. The static spectrum of Xz acquired at 19.5 T reveals the presence of a small quantity of **Xh** that arises from the uptake of moisture (Figure 1). This is supported by the ³⁵Cl static pattern of Xh that has a sharp set of discontinuities at the same position. Evidence of an impurity is also observed in both the ³⁵Cl static and MAS spectra of **Xy** acquired at 18.8 T (Figure 2), which indicates that the sample converts to Xh over time (samples were shipped from Windsor to Tallahassee in tightly sealed containers; nonetheless, some interconversion occurred, perhaps due to the accumulation of trace water on the glass surfaces). Overlays of simulated static and MAS spectra of Xy and Xh clearly indicate the presence of Xh as a minor component; the static spectrum is more reliable for deconvolution and measurement of integrated intensities,⁹ with patterns corresponding to Xy and Xh having a ratio of integrated intensities of 6.5:1.

3.3. Cocrystals. *3.3.1. Mechanochemical Synthesis.* We synthesized three novel cocrystals mechanochemically by NG of X with coformers Bzc, Bza, and Mal, in the appropriate stoichiometric ratios, yielding cocrystals XBzc, XBza, and X₂Mal, respectively. We attempted each synthesis with varying solid forms of X used in the ball milling reactions (Xa, Xz, Xh, and Xx), and each led to the same product. Neat grinding yielded pure white, crystalline, powdered products, which were distinguished from the mixtures of educts *via* a combination of PXRD (Figures S3–S5), ¹³C SSNMR (Figure S15), and ³⁵Cl

	contact type ^a	H…Cl ⁻ distance ^b (Å)	$X \cdots Cl^-$ distance ^c	X–H…Cl [–] angle ^d (deg.)	$C_{\rm Q}$ (MHz)	$\eta_{\rm Q}$	$\delta_{ m iso}~(m ppm)$
Xh	HOH…Cl⁻	2.178	3.111	177.9	3.80	0.53	80
	HOH…Cl-	2.211	3.121	165.2			
	RR′NH…Cl⁻	2.253	3.211	167.2			
Xa	$RR'NH^+\cdots Cl^-$	2.155	3.090	159.3	-3.56	0.63	141
	RR′NH…Cl⁻	2.237	3.147	154.7			
Xx	RR′NH…Cl⁻	2.161	3.131	168.4	-4.72	0.50	76
	$RR'NH^+\cdots Cl^-$	2.354	3.186	143.1			
Ху	RR′NH…Cl⁻	2.097	3.081	177.6	-6.08	0.17	95
	$RR'NH^+\cdots Cl^-$	2.245	3.091	144.6			
Xz	$RR'NH^+\cdots Cl^-$	2.138	3.083	161.0	-3.40	0.99	87
	$RR'NH\cdots Cl^{-}$	2.209	3.121	154.4			
XBzc	ROOH…Cl⁻	2.052	2.999	172.3	-5.22	0.68	112
	RR′NH…Cl⁻	2.102	3.076	171.1			
XBza	RR′NH…Cl⁻	2.176	3.135	165.2	-5.06	0.60	89
	$RNH_2 \cdots Cl^-$	2.390	3.352	174.4			
	$RNH_2 \cdots Cl^-$	2.405	3.342	164.9			
X ₂ Mal (Cl1)	ROOH…Cl⁻	2.040	2.988	171.0	3.98	0.35	109
	RR′NH…Cl⁻	2.118	3.083	168.5			
	$RR'NH^+\cdots Cl^-$	2.285	3.159	149.2			
X ₂ Mal (Cl2)	ROOH…Cl⁻	2.050	3.000	175.7	4.93	0.91	144
	RR′NH…Cl⁻	2.149	3.092	161.8			
	$RR'NH^+\cdots Cl^-$	2.406	3.275	149.3			

Table 3. List of H…Cl⁻ Contacts and Calculated ³⁵Cl SSNMR Parameters for Xylazine HCl Polymorphs, Hydrates, and Cocrystals

^{*a*}Indicates the functional group involved in the H···Cl⁻ bond (*i.e.*, RR'NH₂⁺···Cl⁻ signifies a positively charged secondary amine contact, RR'NH₂^{-/·}Cl⁻ signifies a neutral secondary amine contact, and HOH···Cl⁻ indicates a water molecule contact). ^{*b*}The shortest H···Cl⁻ hydrogen bonds (<2.6 Å), as determined *via* energy minimization and geometry optimization with RPBE-D2* DFT plane-wave calculations. ^{*c*}The distance between the chloride ion and the hydrogen-bond donor atom (X = N, O). ^{*d*}Angle between the hydrogen-bond donor atom (X = N, O), the hydrogen atom, and the chloride ion.

SSNMR (Figure 3). Mechanochemical syntheses using other coformers were also attempted; however, these ball milling experiments resulted in physical mixtures of educts rather than new cocrystals.

3.3.2. Thermal Analysis. DSC thermograms and TGA curves for the cocrystals of X are found in Figures S12 and S13. The DSC thermograms for XBzc and X₂Mal feature single endothermic peaks, likely corresponding to the melting of the cocrystals, with subsequent mass loss in the TGA plot for X₂Mal attributable to the decomposition of malonic acid at 135 °C. The DSC thermogram for XBza features three endothermic peaks, likely corresponding to the decomposition of the cocrystal and the release of HCl, followed by the melting of benzamide (*ca.* 125 °C) and freebase xylazine (*ca.* 140 °C).⁹⁷

3.3.3. Crystal Structures. Following the identification of unique products by PXRD and SSNMR, a small portion of each product was dissolved and recrystallized by slow solvent evaporation, yielding crystals suitable for analysis by SCXRD (Figures S19–S21). Predicted powder patterns based on structures determined from SCXRD data matched initial experimental PXRD data, confirming the formation of the cocrystals **XBzc**, **XBza**, and **X**₂**Mal** upon initial grinding. These structures were solved in the monoclinic C2/c (no. 15, Z = 8, Z' = 1), triclinic $P\overline{1}$ (no. 2, Z = 2, Z' = 1), and orthorhombic $P2_12_12_1$ (no. 19, Z = 4, Z' = 1) space groups, respectively. The asymmetric units of **XBzc** and **XBza** feature a single API molecule, one crystallographically distinct chloride ion, and one coformer molecule. The crystal structure of **X**₂**Mal** features two crystallographically distinct chloride ions. Crystallographic

parameters and additional details of all single-crystal structures are found in Tables S12 and S13.

3.3.4. ${}^{1}H \rightarrow {}^{13}C$ SSNMR Spectra. ${}^{13}C$ CP/MAS spectra for the cocrystals have similar arrays of peak groupings as observed for the other solid forms of X (Figure S15), with the exception of the shifts corresponding to the coformer molecules. Peaks corresponding to the carboxylic acid or amide carbon atoms of the coformers appear at ca. 175-180 ppm, with distinct shifts for each cocrystal. This suggests that there is no transfer of protons between the charged molecules of X and the coformers. The large number of overlapping peaks observed in the spectrum of X₂Mal is consistent with a crystal structure that has two crystallographically distinct molecules of X. None of the spectra show evidence of unreacted educts, although there is some indication of an impurity phase in the spectrum of X₂Mal. As with the case of the monohydrate and anhydrous polymorphs of X, DFT calculations of ¹³C chemical shifts are provided in Figure S16.

3.3.5. ³⁵Cl SSNMR Spectra. The ³⁵Cl NMR spectra of the cocrystals (Figure 3) reveal CT patterns that are distinct from those of all other forms of X, as well as one another, and do not indicate the presence of educts or impurities. The ³⁵Cl NMR spectra of X₂Mal reveal two overlapping patterns, indicating the presence of two crystallographically distinct chlorine sites (*i.e.*, Cl1 and Cl2). The two patterns can be assigned unambiguously *via* the use of DFT calculations of the ³⁵Cl EFG tensors, in which Cl1 corresponds to the narrow CT pattern ($C_Q = 4.28$; $\eta_Q = 0.30$), and Cl2 corresponds to the broad CT pattern ($C_Q = 4.73$; $\eta_Q = 0.97$). For the high-field MAS spectrum, the integrated intensity of Cl1 is slightly greater than that of Cl2, with a ratio of 1.1:1.0 (N.B. the



Figure 4. Relationships between principal components of the ³⁵Cl EFG tensors that have been measured experimentally and determined with DFT-D2* calculations. Results are shown for unrefined structures obtained by X-ray diffraction (left), as well as structures refined at the RPBE-D2* level (right). Dotted lines represent perfect agreement between calculated and experimental values. Γ_{RMS} and N denote the root-mean-square EFG distance and the number of chloride ion sites, respectively.

spectrum was acquired using a RAPT-echo sequence with CTselective pulses, which results in uniform nutations of individual isochromats, ensuring that integrated intensities can be regarded as quantitative. Additionally, since values of T_2^{eff} for chloride ions are typically on the order of 2–20 ms, the interpulse delay of 10 μ s used here has limited impact on integrated intensities).

3.4. Plane-Wave DFT Calculations. Dispersion-corrected plane-wave DFT-D2* calculations can be used to obtain geometry-optimized structures for periodic systems, as well as to calculate NMR interaction tensors based on experimentally determined or computationally refined crystal structures (Table 3; a comparable analysis for the unrefined structures is provided in Table S14). Herein, the relationships between experimentally measured ³⁵Cl EFG tensors, which are very sensitive to chloride ion hydrogen-bonding arrangements, are used as metrics to assess the validity of structures. Calculations are performed on models based on PXRD or SCXRD structures, as well as model structures that have been refined with dispersion-corrected DFT-D2* calculations. The DFT-D2* calculations have been parameterized in previous work from our laboratory to provide accurate placements of hydrogen atoms, based on a comparison of experimentally measured and theoretically derived EFG and CS tensors for several nuclides.^{89–91} Since there are relatively straightforward relationships between the local structures of the chloride ions and the ³⁵Cl EFG tensor parameters and orientations (vide infra),^{30,31,36,54} we explore this further for all of the systems described herein.

The experimental ³⁵Cl EFG and CS tensors are compared with those obtained from plane-wave DFT calculations using model structures based on experimentally determined structures (SCXRD or Rietveld refinement of PXRD data), and DFT-D2* refinements involving the repositioning of all atoms within a fixed unit cell (Table S15).^{89–91} The improved quality of each crystal structure following geometry optimization at the RPBE-D2* level is indicated by the following criteria: (i) the reduction of the static lattice energies, (ii) the reduction of the RMS Cartesian forces on all atoms; and (iii) the reduction of the ^{35}Cl EFG distances, Γ_{EFG} , for each chloride ion. Comparison of the experimental and calculated ³⁵Cl EFG tensors (Figure 4) for the XRD-derived structures reveals very poor agreement, with an RMS Γ_{EFG} of 1.58 MHz for the nine chloride ions, whereas improved agreement is observed for the refined structures, with an RMS Γ_{EFG} of 0.27

MHz. Calculation on our refined crystal structure of Xh results in excellent agreement with experiment ($\Gamma_{\rm EFG} = 0.26$ MHz), whereas calculations on five previously reported structures do not,^{60,61} even though all geometry-optimized structures of Xh result in nearly identical static lattice energies (Table S16). The agreement between experimental and calculated chlorine CS tensors is more ambiguous, due to the fact that the CS tensor parameters and Euler angles are extracted from spectra dominated by contributions from the SOQI, and are therefore associated with higher uncertainties; these parameters are not discussed further.

Thermal effects on calculated ³⁵Cl EFG tensors have been discussed previously.⁹¹ In general, the effect of small lattice expansions/contractions associated with changes in temperature have only small effects on calculated EFG tensors. For example, the calculated values of $C_{\rm Q}$ for the chloride ion in L-histidine HCl·H₂O differ by only 120 kHz for DFT-D2* geometry-optimized structural models based on SCXRD structures obtained at 298 and 100 K. However, in rare instances, motions of highly mobile moieties, such as NH₄⁺ ions, can impact both calculated and experimental EFG tensors; in such cases, these effects can be modeled using *ab initio* molecular dynamics.⁵⁴

The types and arrangements of hydrogen bonds around chloride ions are correlated to the ³⁵Cl EFG tensors (Table 3).^{30,31,36,54,57} Hydrogen bonds with the largest influence on the ³⁵Cl EFG tensors are those in which the H…Cl⁻ distance is less than ca. 2.2 Å, which we define here as short contacts, whereas all interactions up to ca. 2.6 Å are referred to as long contacts.¹⁰⁰⁻¹⁰² When systems have a single short contact, DFT calculations invariably show that the sign of C_0 is negative (*i.e.*, the sign of V_{33} is positive, since $Q(^{35}Cl) = -8.165 \text{ fm}^2)$,¹⁰³ and the largest principal component of the EFG tensor, V_{33} , is oriented in the general direction of the short contact. When systems have two short contacts, calculations indicate that the sign of $C_{\rm O}$ is positive, and the negative principal component V_{33} is oriented approximately perpendicular to the plane formed by the two short contacts (*i.e.*, EFGs are positive in the directions of the two short contacts). However, in situations in which the two short contacts are aligned in a linear (or nearly linear) spatial arrangement, V₃₃ is directed along the hydrogen-bonding axes, and the magnitude of C_Q is predicted to be negative.³⁸ In contrast, for systems that do not feature short contacts (i.e., all hydrogen-bonding distances are greater than *ca.* 2.2 Å), the



Figure 5. ³⁵Cl EFG tensor orientations for the different forms of X. The short (<2.2 Å) H…Cl contacts are shown in red. Hydrogen bonds (2.2–2.6 Å) are shown in green. Yellow vectors represent the orientations of the principal components of the EFG tensor (V_{11} , V_{22} , and V_{33}), where one vector is oriented perpendicular to the page.

relationships between EFG tensors and molecular-level structure are more complicated and vary from system to system, and generally have C_Q values of small magnitude.^{31,36} As such, it is of interest to explore the relationships between the EFG tensor parameters, tensor orientations, and the chloride ion environments for the solid forms of X, as such information may be useful in future NMR crystallographic investigations of novel forms of APIs without crystal structures.

The calculated ³⁵Cl EFG tensor orientations for the refined structures of the solid forms of X are shown in Figure 5 (N.B.: the EFG tensor is centrosymmetric; however, we have chosen to use three yellow vectors to simplify the diagrams). For the four anhydrous polymorphs of X, the chloride ion resides in an environment featuring a single short contact and one additional long contact. In each case, calculations show that the value of C_Q is negative. Xa, Xx, and Xz feature an N3–C4– N5-C6 torsion angle near 180°, and V_{33} resides within the plane formed by the short and long contacts due to the spatial proximity of these two hydrogen-bond donating sites. In contrast, Xy features a torsion angle near 0° , and V_{33} is oriented in the direction of the short contact. Similarly, for **XBzc** and **XBza**, DFT calculations show that the value of C_0 is negative and V_{33} is oriented in the direction of the shortest contact, even when a second short contact is present, as is the case for XBzc. In contrast, the structure of Xh features two

short contacts with water molecules, and calculations show that the sign of C_Q is positive, with V_{33} oriented approximately perpendicular to the plane formed by the chloride ion and the two short contacts. Calculations on the two chloride ions in X_2 Mal also show that their values of C_Q are positive, and V_{33} components are oriented approximately perpendicular to the plane formed by the chloride ion and the two short contacts in each case.

In summary, the orientations of the ³⁵Cl EFG tensors are largely consistent with our previous observations on organic chloride salts. It is expected that this expanding body of data will be useful for gaining insight into crystal structures in the absence of structural data obtained from diffraction studies, and may even serve as the groundwork for future applications in NMR-guided crystal structure prediction, involving the EFG tensors as a primary metric, and possibly even artificial intelligence and/or machine learning, as has been recently explored with chemical shifts.^{104–106}

4. CONCLUSIONS

We have determined optimized experimental conditions for synthesizing three previously reported anhydrous polymorphs, a monohydrate, and a novel anhydrous polymorph of xylazine HCl, as well as three new cocrystals with X as a component and carboxylic acids and an amide as coformers, all with high yields and rapidity. We characterized the microcrystalline samples with thermal analysis, PXRD, and multinuclear SSNMR spectroscopy, and determined crystal structures for the novel anhydrous polymorph, the hydrate, and three new pharmaceutical cocrystals. Additionally, dispersion-corrected plane-wave DFT-D2* calculations were used to obtain refined crystal structures, from which ³⁵Cl EFG tensors were computed and found to agree with experiment. The combination of ³⁵Cl SSNMR, PXRD, and quantum chemical calculations is invaluable for spectral fingerprinting, examining interconversion of polymorphs and hydrates, optimizing synthetic protocols, making structural assignments, and exploring the relationships between the local chloride ion environments and the ³⁵Cl EFG tensors. These combined methodologies should prove useful for future optimizations of mechanochemical syntheses of both polymorphs and cocrystals, perhaps even allowing for the rational design of new solid forms and aiding in de novo NMR crystallographic crystal structure prediction.

ASSOCIATED CONTENT

3 Supporting Information

The Supporting Information is available free of charge at https://pubs.acs.org/doi/10.1021/acs.cgd.2c01539.

Synthetic conditions; NMR acquisition parameters; summary of computational results; crystallographic information; PXRD; thermal analysis; and additional SSNMR spectra (PDF)

Accession Codes

CCDC 2214351–2214355 contain the supplementary crystallographic data for this paper. These data can be obtained free of charge via www.ccdc.cam.ac.uk/data_request/cif, or by emailing data_request@ccdc.cam.ac.uk, or by contacting The Cambridge Crystallographic Data Centre, 12 Union Road, Cambridge CB2 1EZ, UK; fax: +44 1223 336033.

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L.M.A., A.A.P., and S.T.H. made equal contributions to this manuscript. L.M.A. was responsible for the majority of sample preparations and initial manuscript drafts, A.A.P. was responsible for the design of synthetic procedures and characterization of products, and S.T.H. was responsible for first-principles calculations and final manuscript drafts. All three contributed to sample characterization using solid-state NMR, PXRD, and other methods. Z.T.D. assisted with refining mechanochemical syntheses and obtaining ¹³C CP/MAS spectra. L.K.W. and J.M.R. were responsible for single-crystal XRD experiments and structure determination. E.M.I. and D.A.H. prepared some preliminary samples and collected SSNMR and PXRD data for them. L.M.A., A.A.P., S.T.H., J.M.R., and R.W.S. made significant contributions to the writing and editing of this manuscript

Notes

The authors declare no competing financial interest.

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